

# Enzymatic fluorination of biotin and tetrazine conjugates for pre-targeting approaches to PET imaging.

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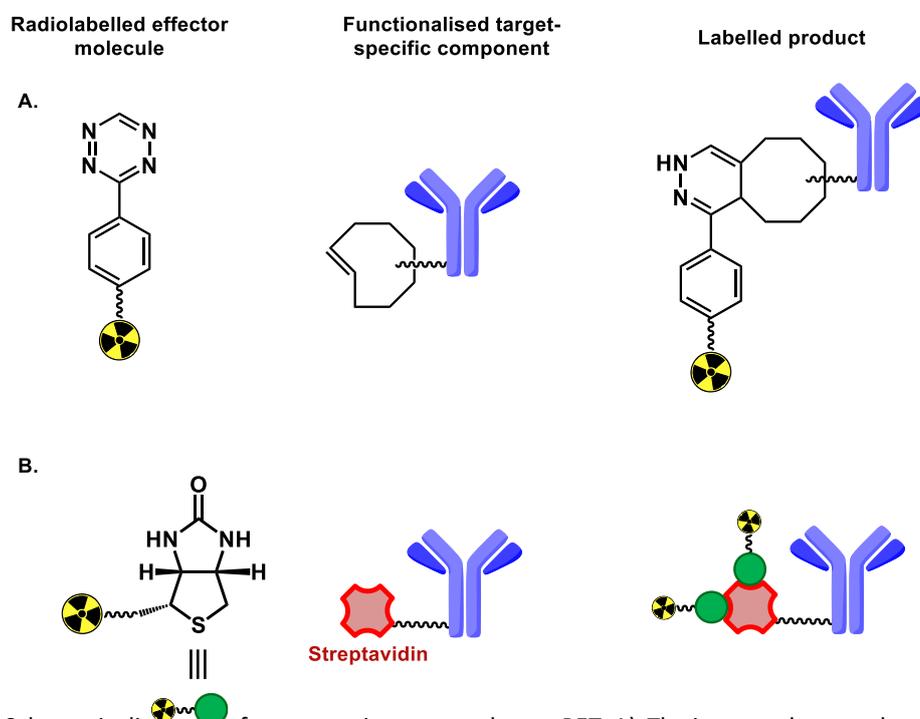
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The use of radiolabelled antibodies and antibody-derived recombinant constructs have shown promise for both imaging and therapeutic use. In this context, the biotin-avidin/streptavidin pairing along with the inverse electron demand Diels–Alder reaction (IEDDA) have found application in pre-targeting approaches for positron emission tomography (PET). This study reports the fluorinase enzyme mediated transhalogenation (CIDA substrates to FDA products) of two antibody pre-targeting tools, a FDA-PEG-tetrazine and a [<sup>18</sup>F]FDA-PEG-biotin and each is assessed for their compatibility towards IEDDA ligation to *trans*-cyclooctene or affinity to avidin. A protocol was developed to avoid radiolytically promoted oxidation of biotin during the synthesis of [<sup>18</sup>F]FDA-PEG-biotin. The study adds to the repertoire of conjugates for use in fluorinase catalysed radio-synthesis for PET and shows that the fluorinase will accept a wide range of CIDA substrates tethered at C-2 of the adenine ring with a pegylated cargo. The method is exceptional as the nucleophilic reaction with [<sup>18</sup>F]fluoride takes place in water at neutral pH and at ambient temperature.

## Introduction

Positron emission tomography (PET) has emerged as a valuable tool in clinical imaging due to its high sensitivity and complementarity to other *in vivo* clinical imaging modalities, such as computerised tomography (CT) and magnetic resonance imaging (MRI).<sup>[1]</sup> [<sup>18</sup>F]Fluoride is the most commonly used radionuclide for PET imaging as it possesses a relatively long half-life ( $t_{1/2} = 109.8$  min) for a clinical positron emitter and image resolution is enhanced as it has a short positron range (2.3 mm in water).

A recent drive to improve imaging performance by PET has led to a shift from the direct labelling of traditional low molecular-weight and rapidly circulating (hours) radiotracers, which often suffer non-specific binding and poor localisation *in vivo*, to the use of large but very high affinity proteins such as monoclonal antibodies or their fragments, which circulate much more slowly (days).<sup>[2]</sup> Such proteins cannot be usefully labelled by fluorine-18 at point of injection as the isotope half-life is inconsistent with the pre-incubation time required for their location at the target. Instead, a pre-targeted approach is required where the antibody is administered in its unlabelled form, but with a reactive motif attached, and then several hours to days later, a low molecular weight <sup>18</sup>F-labelled reactive partner is administered which circulates rapidly and covalently modifies the *in situ* antibody. Such strategies have begun to exploit bioorthogonal click reactions<sup>[3]</sup> and make use, for example, of strain promoted alkyne–azide cycloaddition (SPAAC) or inverse electron demand Diels–Alder reactions (IEDDAs), without the need for catalysis.<sup>[4]</sup> IEDDA based methods in particular involve complimentary tetrazine and trans-cyclooctene (TCO) motifs which combine remarkably rapidly (reaction rates up to  $k_2$  2000  $M^{-1} s^{-1}$  in 9:1 methanol/water),<sup>[4d]</sup> allowing for protein modification at low radiotracer concentrations.



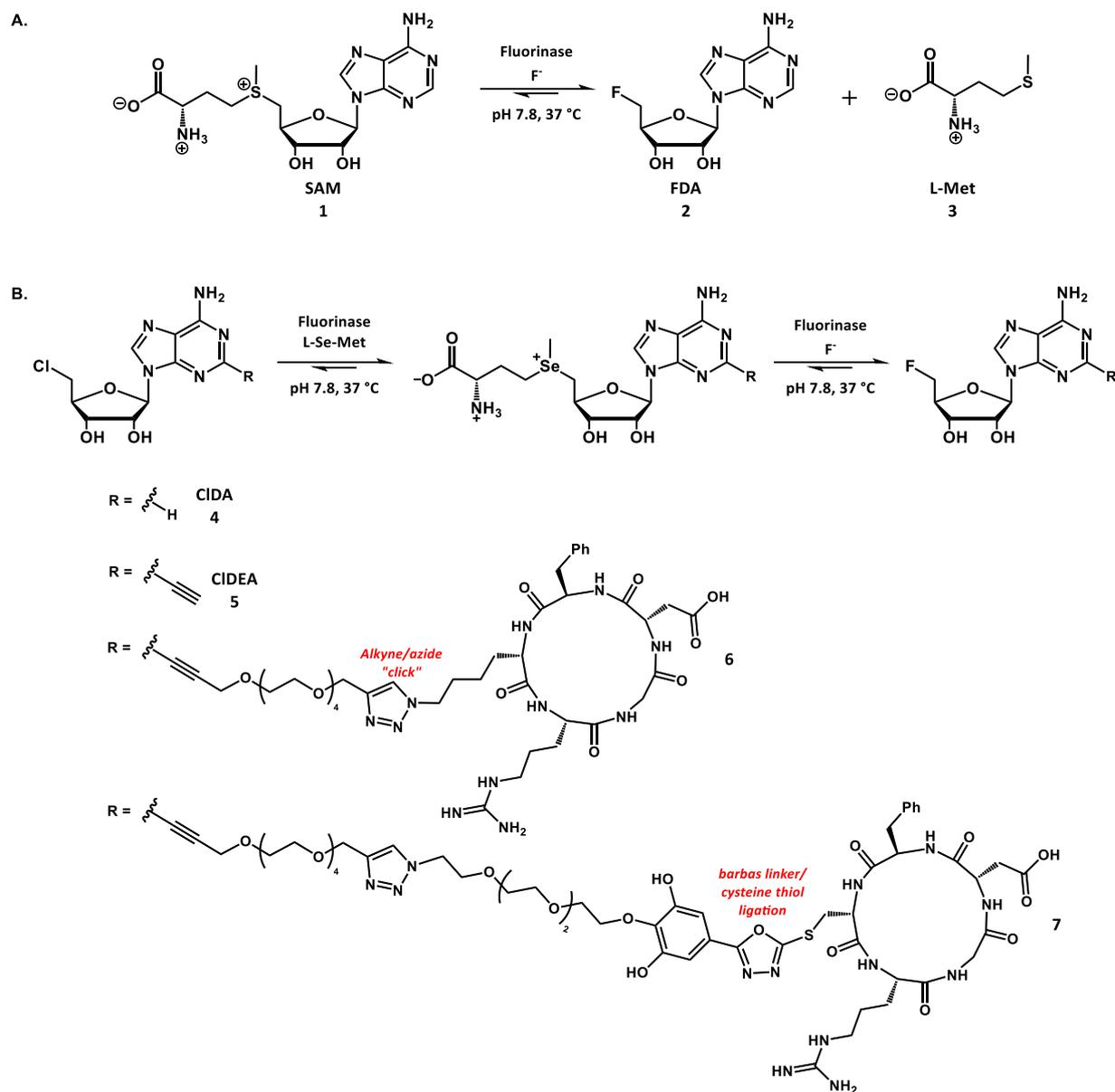
**Scheme 1.** Schematic diagrams of pre-targeting approaches to PET. A) The inverse electron demand Diels–Alder (IEDDA) bioorthogonal click reaction. B) The (+)-biotin/(strep)avidin affinity pairing.

In this context, the initial approach has been to fluorinate the TCO motif as direct nucleophilic fluorination of the reactive tetrazine motif is challenging due to decomposition.<sup>[5]</sup> Thus efficient

methods for radiolabelling tetrazines with fluorine-18 are desirable.<sup>[6]</sup> In this regard an Al<sup>[18F]</sup>NOTA-labelled tetrazine tracer was successfully utilised in a pre-targeting PET study, expressing BxPC3 pancreatic cancer xenografts in mice.<sup>[4c]</sup> There remains room to explore other <sup>18</sup>F-labelled tetrazine strategies and in this study we have investigated enzymatic fluorination in water/buffer under ambient conditions.

Similarly, pre-targeting strategies have been designed around the (+)-biotin/(strep)avidin affinity pairing.<sup>[7]</sup> The interaction is among the strongest non-covalent interactions ( $K_d = 1 \times 10^{-15}$  M) known and is unaffected by extremes of pH and temperature. Initial approaches to multistep imaging involved the use of unlabelled antibodies conjugated to (+)-biotin, followed by the administration of radiolabelled (strep)avidin.<sup>[8]</sup> Alternatively, a more efficient pre-targeting approach involves the administration of large (strep)avidin-antibody conjugates, followed by a rapidly circulating, low molecular weight [<sup>18</sup>F]biotin derivative.<sup>[9]</sup> There has been concern when employing this strategy regarding the role of endogenous biotin competing with the radiolabelled chaser *in vivo*, though this has been successfully addressed in mice models through a biotin-deficient diet.<sup>[10]</sup>

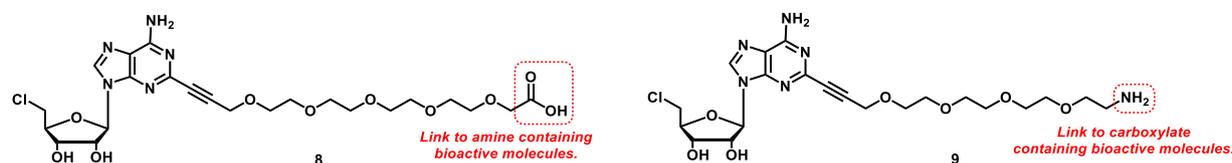
Radiolabelling of (+)-biotin has generally been achieved through the pre-synthesis and then coupling of <sup>18</sup>F-labelled prosthetic groups, typically [<sup>18</sup>F]4-fluorobenzylamine<sup>[7b]</sup>, p-<sup>[18</sup>F]fluorobenzoic acid<sup>[10b]</sup> and [<sup>18</sup>F]FDG<sup>[11]</sup>. The approach involves several steps and requires efficient fluorination strategies. Direct labelling approaches, such as the nucleophilic fluorination of a (+)-biotin-mesylate precursor are more appealing.<sup>[12]</sup> We have been exploring an even more direct approach involving enzymatic fluorination, which can be accomplished without drying the [<sup>18</sup>F]fluoride which is generated in a cyclotron in [<sup>18</sup>O]water, prior to nucleophilic chemistry. Such an approach requires minimal manipulation of the radioisotope. A similarly direct approach has been explored with boronaryl functionalized (+)-biotin,<sup>[13]</sup> however in order to avoid deboronation during the radiolabelling protocol, a low pH is required.



**Scheme 2.** A) The native reaction catalyzed by the fluorinase enzyme. B) Fluorinase mediated transhalogenation reaction of C-2 modified substrates, **6** and **7** exemplify previously achieved alkyne/azide "click"<sup>[14]</sup> and barbas linker/thiol-ligation<sup>[15]</sup> bioconjugation strategies.

The fluorinase enzyme catalyses the conversion of S-adenosyl-L-methionine (AdoMet) and fluoride ion to generate 5'-fluoro-5'-deoxyadenosine (5'-FDA, Scheme 2).<sup>[16]</sup> It also catalyses the nucleophilic displacement of chloride by L-methionine in the reverse direction to generate AdoMet,<sup>[17]</sup> and if fluoride is present in this reaction 5'-FDA is formed in a sequential step, such that an overall transhalogenation reaction occurs. Moreover, the replacement of AdoMet with AdoSeMet increases the overall transhalogenation efficiency, as selenium is a better nucleophile and leaving group than sulfur (Scheme 2). The fluorinase displays a high degree of substrate specificity, however it can tolerate an acetylene moiety at the C2 position on the adenine base of 5'-CIDA, additionally a pegylated cargo can also be tethered to the terminus of this acetylene. In this way the fluorinase will catalyse 'last-step' aqueous [<sup>18</sup>F]fluorination of 5'-CIDA-PEG-peptides.<sup>[14]</sup> To date, methodologies to

permit bioconjugation of RGD peptides to the fluorinase binding 5'-CIDA moiety either through 'click' based chemistry (*via* azide functionalized peptides)<sup>[14, 18]</sup> or utilising a modified Barbas bioconjugation strategy (*via* cysteine containing peptides)<sup>[15]</sup> have been developed (Scheme 2). Recently, the fluorinase enzyme has also become the target of engineering *via* directed evolution, in an effort to achieve higher radiochemical conversions (RCC) and broaden the structural diversity of substrates.<sup>[19]</sup>



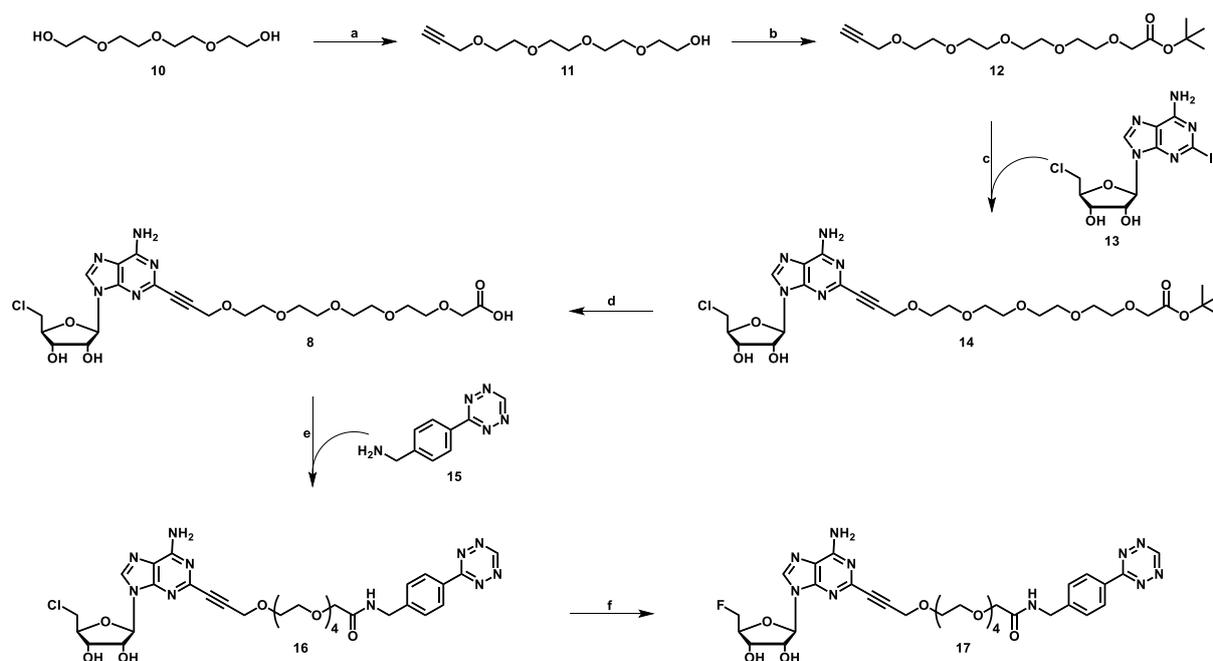
**Scheme 3.** Design of fluorinase binding CIDA-PEG-acid **8** and CIDA-PEG-amine **9** constructs for amide bond ligation to biologically relevant molecules.

In order to demonstrate a wider array of conjugation methods (employing amide linkage *via* CIDA-PEG-acid **8** and CIDA-PEG-amine **9**, Scheme 3) compatible with fluorinase labelling of higher molecular-weight proteins, we now report the fluorinase mediated transhalogenation of CIDA-PEG-tetrazine **16** to FDA-PEG-tetrazine **17** demonstrating compatibility with the tetrazine ring under ambient, aqueous conditions. Similarly CIDA-PEG-biotin **26** was used in a proof of principle study to deliver a biotin radiotracer, [<sup>18</sup>F]**27**, as a tool for radiolabelling pre-targeted (strept)avidin conjugated antibodies.

## Results and Discussion

### Synthesis of CIDA-PEG-acid **8** and CIDA-PEG-tetrazine **16**

A three step approach was employed for the synthesis of both CIDA-PEG-tetrazine **16** and CIDA-PEG-biotin **26**.<sup>[14, 18b]</sup> This involved the synthesis of a fluorinase compatible 5'-chloro-5'-deoxyadenosine attached to an amine (**9**) or carboxylic acid (**8**) functionality through a PEG spacer. Amide coupling to either benzylamino tetrazine **15** or the carboxylate of (+)-biotin **25**, generated tetrazine **16** (Scheme 4) or CIDA-PEG-biotin **26** (Scheme 5) constructs respectively. Briefly, 5'-chlorodeoxy-2-iodo-adenosine **13**, a key intermediate in these syntheses, was prepared from guanosine using a previously established protocol.<sup>[18b]</sup> The synthesis of **16** required that the pegylated linker was functionalized with an alkyne terminus. This was accomplished by first treating tetraethylene glycol **10** with propargyl bromide to provide the monoalkylated PEG derivative **11**, which, after purification, was treated with tert-butyl bromoacetate to afford **12**. This PEG linker serves two purposes; firstly, it aids in the aqueous solubility of the conjugate, an essential requirement for fluorinase catalysis. Secondly, the linker serves to project any functionalized moiety from the surface of the fluorinase and into the bulk solvent.

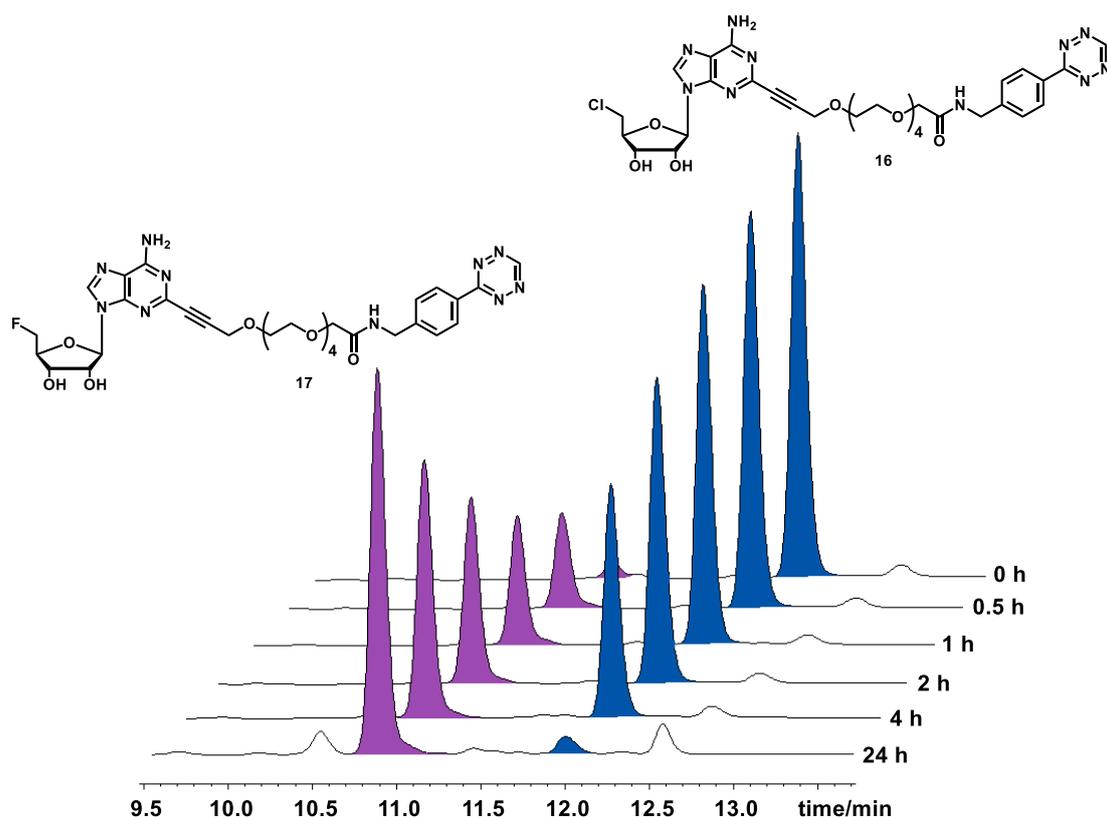


**Scheme 4.** Synthesis of CIDA-PEG-acid **8**, CIDA-PEG-tetrazine **16** and FDA-PEG-tetrazine **17**. Reagents and conditions: a) NaH, propargyl bromide, THF, 0 °C to r.t., overnight; b) NaH, tert-butyl bromoacetate, THF, 0 °C to r.t., overnight; c) Pd<sub>2</sub>(dba)<sub>3</sub>, Et<sub>3</sub>N, CuI, DMF, 80 °C, 24 h; d) TFA, DCM, 0 °C to r.t., 45 min; e) PyBOP, DIPEA, DMF, r.t., 24 h; f) Fluorinase, L-Se-Met, KF, phosphate buffer (pH 7.8), 37 °C, 72 h.

With both **12** and **13** in hand, our attention turned to further assembly *via* Sonogashira cross-coupling. This was accomplished,<sup>[20]</sup> using an excess of alkyne **12** over 5'-chlorodeoxy-2-iodo-adenosine **13** in the Pd<sub>2</sub>(dba)<sub>3</sub> catalysed reaction. The CIDA-PEG-ester **14** was recovered in high purity after chromatography and C18 cartridge purification. CIDA-PEG-ester **14** was treated with TFA to generate **8** as a TFA salt, and this could be used directly in the subsequent coupling reactions, without the need for further purification. Finally, an amide coupling reaction (PyBOP, DIPEA) was employed to attach carboxylic acid **8** to benzylamino tetrazine **15** (Scheme 4). This afforded the CIDA-PEG-tetrazine **16** conjugate in high purity after preparative HPLC.

### Evaluation of CIDA-PEG-tetrazine as a substrate for fluorinase mediated transhalogenation

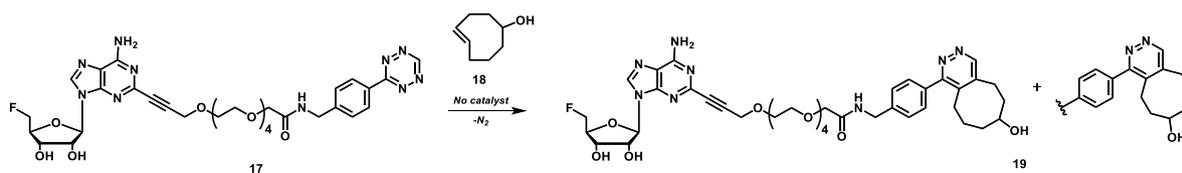
Substrate **16** was incubated with the fluorinase (1 mg mL<sup>-1</sup>, these relatively high amounts are due to the fluorinase having a slow *k*<sub>cat</sub> of 0.03 min<sup>-1</sup>)<sup>[17]</sup> under previously established conditions<sup>[17]</sup> in phosphate buffer at pH 7.8 (see Supporting Information). A time course profile (Figure 1) showed that **16** was an excellent substrate for enzyme catalysed transhalogenation generating FDA-PEG-tetrazine **17**, in ~60% conversion after 4 h, and this offers a new approach to fluorinating tetrazine derivatives. Importantly, no obvious degradation was observed within the initial hours of these experiments (the timeframe required for radiolabeling experiments), indicating that CIDA-PEG-tetrazine and its fluorinated analogue are stable under these conditions. The additional peak observed at 12.5 min is indicative of the intermediate L-Se-Met adduct (which consistently possesses a greater retention time to that of its corresponding CIDA and FDA derivatives).



**Figure 1.** HPLC time course (UV, 254 nm) of the incubation of CIDA-PEG-tetrazine **16**, blue ( $t_R = 11.9$  min), with the fluorinase, L-Se-Met, KF, phosphate buffer (pH 7.8) at 37 °C. Traces show the formation of FDA-PEG-tetrazine **17**, purple ( $t_R = 10.8$  min), and the consumption of CIDA-PEG-tetrazine **16**. For full conditions see the Experimental Section and Supporting Information.

### CIDA-PEG-tetrazine/dienophile IEDDA ligation

With FDA-PEG-tetrazine **17** in hand, it was explored for its ability to participate in IEDDA-initiated ligation. For this study (*E*)-cyclooct-4-enol **18** was chosen as a coupling partner. Thus, a solution (40  $\mu$ l, 50 mM) of (*E*)-cyclooct-4-enol in DMSO was added to an aqueous solution of **17**, and the mixture was incubated at room temperature for 2 min. An aliquot was then subjected to HPLC and HRMS analysis of the new product isolated was consistent with the formation of the fully aromatised ligated product **19** (Scheme 5).



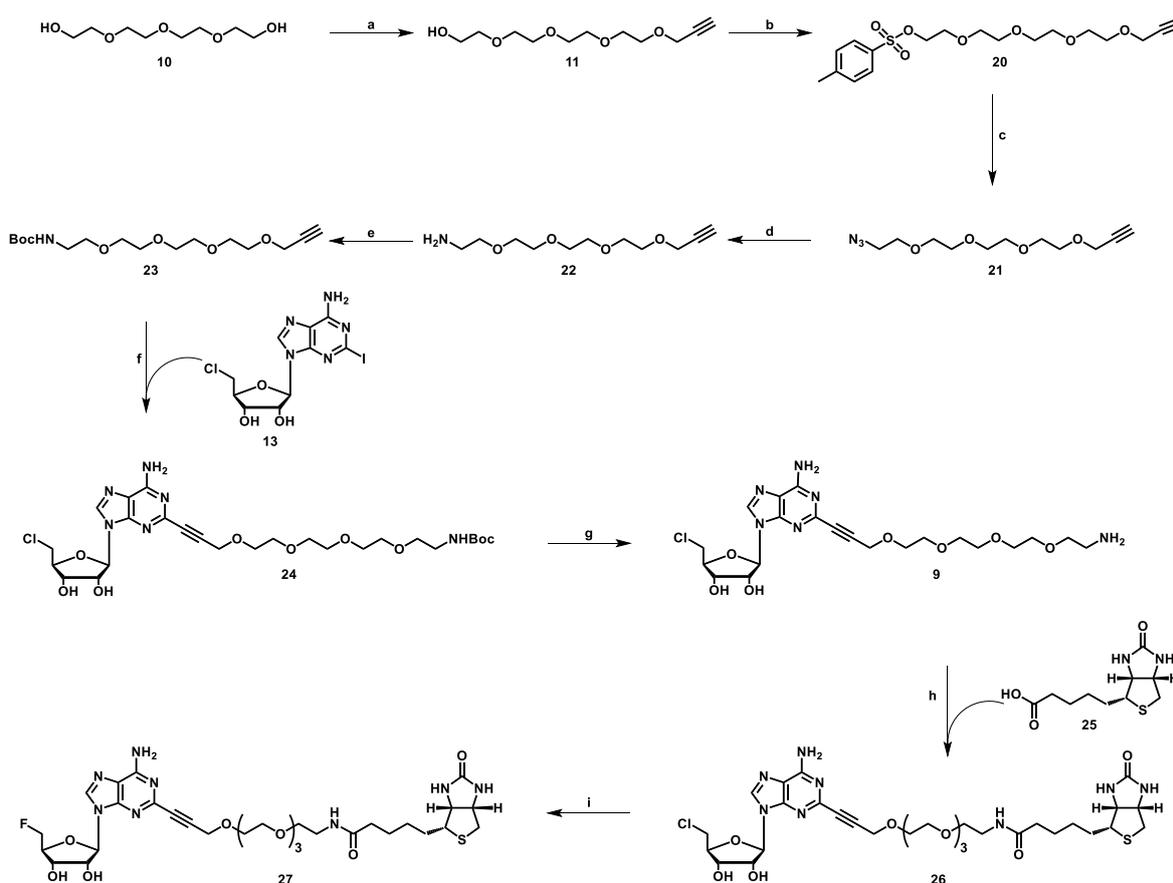
**Scheme 5.** IEDDA initiated conjugation between **17** and (*E*)-cyclooct-4-enol **18** (identity of fully aromatised ligation product **19** was confirmed by HRMS after isolation by HPLC).

## Last step enzymatic <sup>18</sup>F labelling of **16** to [<sup>18</sup>F]FDA-PEG-tetrazine

An enzymatic transhalogenation in the presence of [<sup>18</sup>F]fluoride was investigated with **16** and its conversion into [<sup>18</sup>F]FDA-PEG-tetrazine ([<sup>18</sup>F]**17**) was clearly demonstrated against a product reference. Using the previously established protocol,<sup>[15]</sup> [<sup>18</sup>F]**17** was generated with an approximate 10% radiochemical conversion (decay uncorrected) after a 30 min incubation with the fluorinase (Figure S46). Isolation of [<sup>18</sup>F]**17** by semi-prep HPLC afforded the radiolabelled product with two minor radiolabelled impurities (Figure S47). This process requires optimisation for pre-clinical dosing, however it is clear that the tetrazine moiety is compatible with the fluorinase radiolabelling protocol.

## Synthesis of CIDA-PEG-amine **9** and CIDA-PEG-biotin **26**

The synthesis of CIDA-PEG-biotin **26** followed a similar strategy to that used for CIDA-PEG-tetrazine **16**. Firstly, a pegylated linker was required with alkyne and amine termini. For this alcohol **11** was treated with tosyl chloride and then **20** was subjected to nucleophilic displacement with sodium azide to generate azide **21**. A Staudinger reduction of this azide afforded free amine **22**, which was then Boc protected to generate **23**.

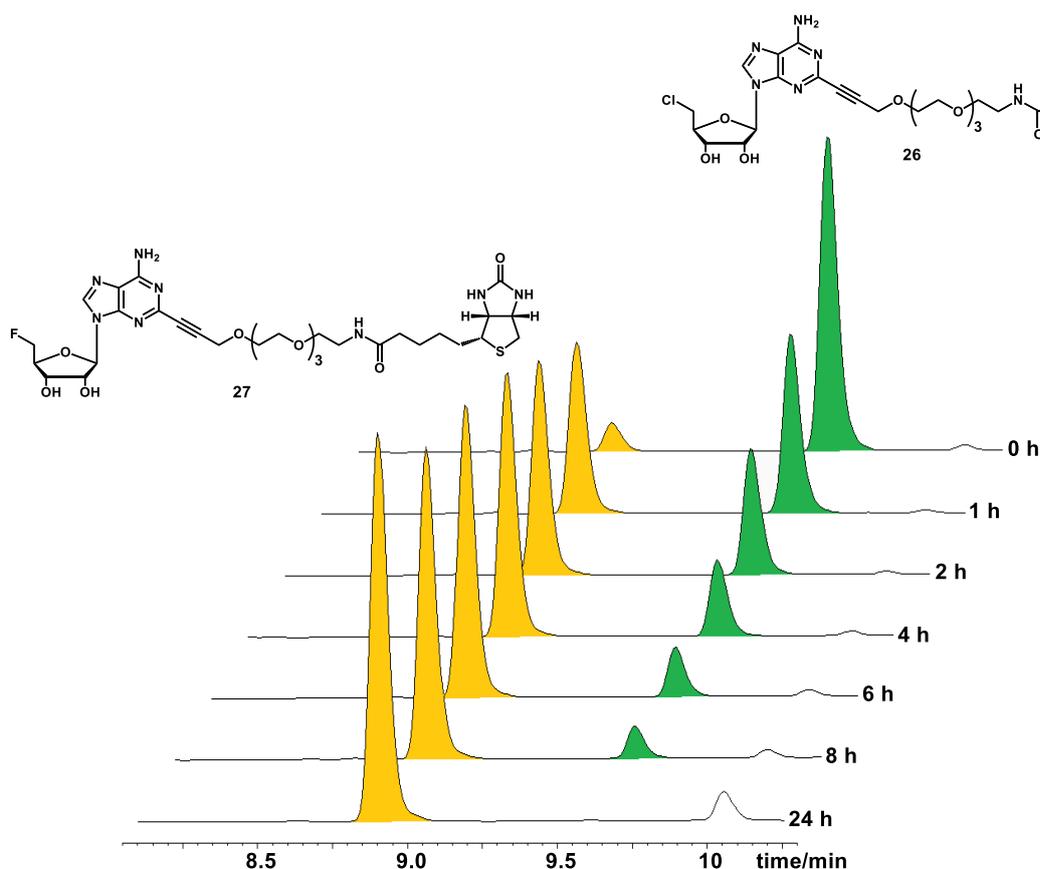


**Scheme 6.** Synthesis of CIDA-PEG-amine **9**, CIDA-PEG-biotin **26** and FDA-PEG-biotin **27**. Reagents and conditions: a) NaH, propargyl bromide, THF, 0 °C to r.t., overnight; b) Tosyl chloride, Et<sub>3</sub>N, THF, r.t., 20 h; c) NaN<sub>3</sub>, DMF, 50 °C, 18 h; d) PPh<sub>3</sub>, H<sub>2</sub>O, THF, r.t., 48 h; e) (Boc)<sub>2</sub>O, Et<sub>3</sub>N, MeOH; f) Pd<sub>2</sub>(dba)<sub>3</sub>, Et<sub>3</sub>N, CuI, DMF, 80 °C, 24 h; g) TFA, DCM, 0 °C to r.t., 45 min; h) PyBOP, DIPEA, DMF, r.t., 24h; i) Fluorinase, L-Se-Met, KF, phosphate buffer (pH 7.8), 37 °C, 48 h.

A Sonogashira cross-coupling was carried out with an excess of alkyne **23** over 5'-chlorodeoxy-2-iodoadenosine **13** using the previously described protocol to generate CIDA-PEG-protected amine **24**, after HPLC purification, in high purity. Deprotection of **24** with TFA led to free amine **9** (as a TFA salt) which was used directly in subsequent coupling reactions, without the need for further purification. As with carboxylic acid **8**, amine **9** offers a complementary tool for the fluorinase [<sup>18</sup>F]-labelling, for conjugation to the C-termini of bioactive peptides. Amide coupling (PyBOP, DIPEA) allowed preparation of the CIDA-PEG-biotin conjugate **26** in good yield and high purity after HPLC purification.

### Evaluation of CIDA-PEG-biotin as a substrate for fluorinase mediated transhalogenation

As a prelude to [<sup>18</sup>F]-radiolabelling a 'cold' transhalogenation reaction with CIDA-PEG-biotin **26** was explored with the fluorinase. Accordingly, substrate **26** was incubated with the fluorinase (1 mg mL<sup>-1</sup>) L-Se-Met (75 μM) and KF (50 mM) in phosphate buffer, at pH 7.8 (see Supporting Information). An analytical time course profile revealed chloride **26** to be an excellent substrate for fluorinase mediated transhalogenation generating FDA-PEG-biotin **27**, with an almost 80% conversion after 4 h. No reaction was observed in the absence of the fluorinase and the water solubility of **26** circumvented any requirement to add DMSO. A significant advantage in such reactions, as adding DMSO can complicate work up by promoting gelling on heat denaturation.

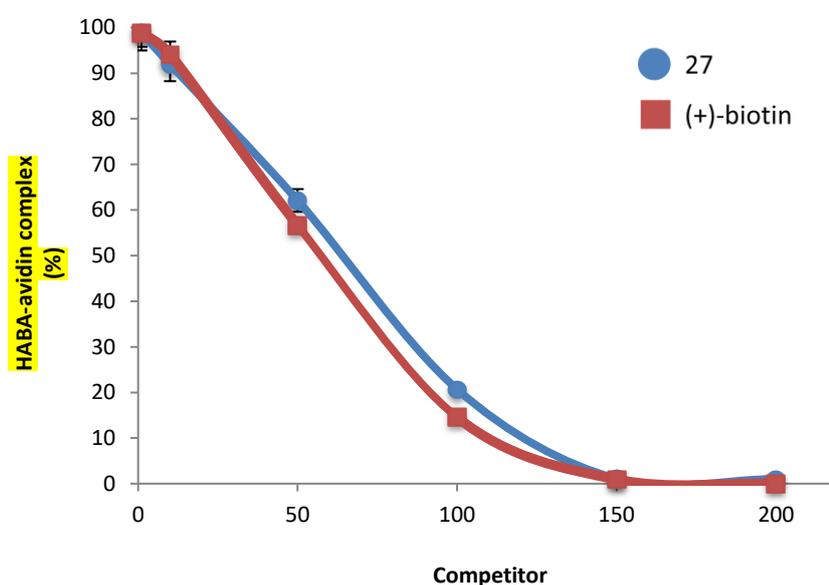


**Figure 2.** HPLC time course (UV, 254 nm) of the incubation of CIDA-PEG-biotin **26**, green ( $t_R = 9.5$  min), with the fluorinase, L-Se-Met, KF, phosphate buffer (pH 7.8) at 37 °C. Traces show the formation of FDA-PEG-biotin **27**, yellow ( $t_R = 8.9$  min), and the consumption of CIDA-PEG-biotin **26**. For full conditions see the Experimental Section and Supporting Information.

The transhalogenation of **26** was carried out on a scale sufficient for isolation of an analytical sample of FDA-PEG-biotin **27** to confirm its identity by NMR. A suitable sample was also required to evaluate its binding affinity to avidin (see Supporting Information). The reaction was again monitored by HPLC and was terminated by heat precipitation and centrifugation to remove the enzyme. Purification was accomplished *via* semi-prep HPLC to obtain **27** (3mg) in high purity.

### FDA-PEG-biotin/avidin Binding Assay

The avidin binding affinity of the newly synthesised FDA-PEG-biotin **27** was determined using a colorimetric HABA assay. The assay exploits the UV spectral changes that occur upon the binding of 4'-hydroxyazobenzene-2-carboxylic acid (HABA) to avidin. The HABA/avidin interaction is a relatively weak one ( $K_d = 5.8 \times 10^{-6}$  M versus that of (+)-biotin/Avidin;  $K_d = 1 \times 10^{-15}$  M), and as such free (+)-biotin, or indeed **27**, will readily displace the HABA dye and result in a decrease in absorbance at 500nm wavelength. FDA-PEG-biotin **27** was found to inhibit the HABA/avidin complex in a dose dependant manner (Figure 3) and with a similar affinity to that between avidin and (+)-biotin. The PEG spacer and the FDA moiety of the construct do not impede the binding interaction of **27** with avidin, thus [ $^{18}$ F]**27** has good potential as a candidate tracer for streptavidin-conjugated antibody PET studies.

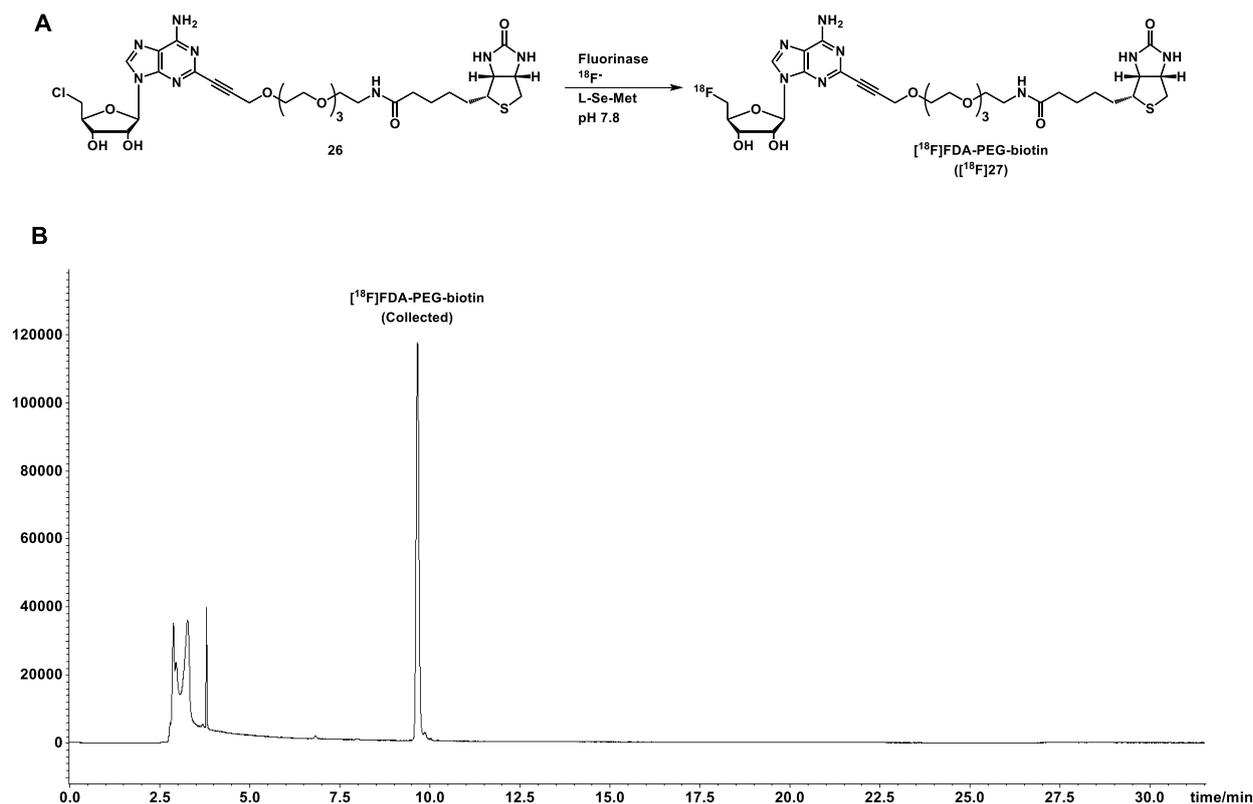


**Figure 3.** Inhibition of the HABA/avidin complex by (+)-biotin (maroon squares) and FDA-PEG-biotin **27** (blue circles). Data are representative of three different experiments.

### Last step enzymatic $^{18}$ F labelling of biotin

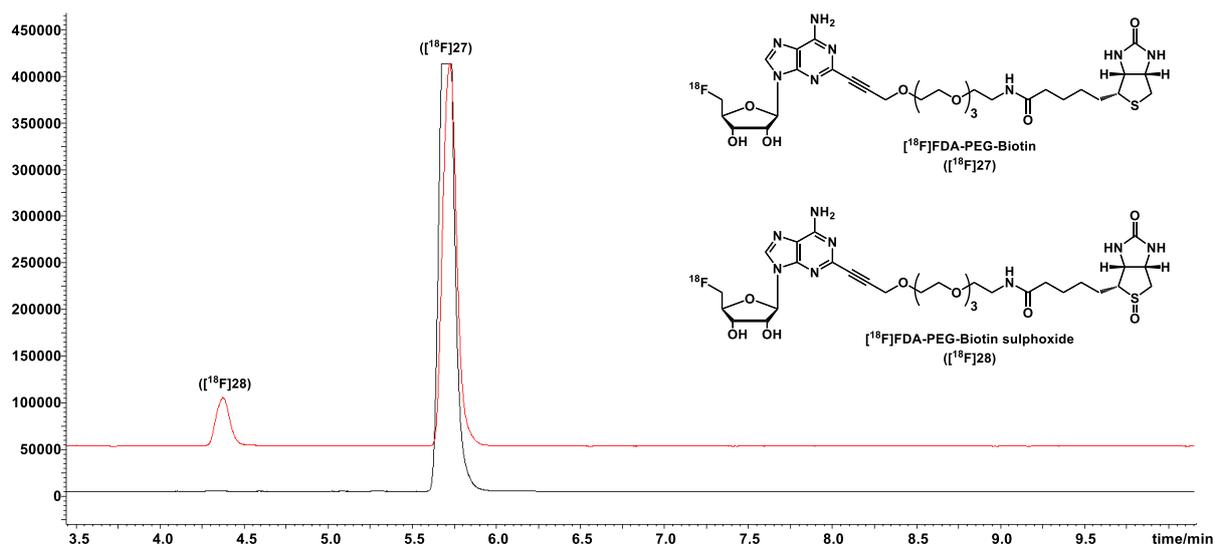
An enzymatic transhalogenation was performed with **26** in the presence of [ $^{18}$ F]fluoride, and its conversion into [ $^{18}$ F]FDA-PEG-biotin ([ $^{18}$ F]**27**) was monitored. For hot labelling an [ $^{18}$ O]aqueous,

solution of [ $^{18}\text{F}$ ]fluoride at GBq levels is generated and then an aliquot (MBq of [ $^{18}\text{F}$ ]fluoride) is added to the biotransformation solution in buffer at pH 7.8 (see Experimental Section). The concentrations are such that these fluorinase-catalyzed radiochemical reactions are no longer catalytic, as the enzyme is in the micromolar range with [ $^{18}\text{F}$ ]fluoride ion in the pico-molar range. In this manner the stoichiometry is dramatically reversed relative to the cold transhalogenation reactions. During these experiments [ $^{18}\text{F}$ ]**27** was produced with a 30-40% radiochemical conversion after only 30 min incubation with the fluorinase. Figure 4 shows the radiochemical HPLC trace of the supernatant of a reaction mixture at this time point.



**Figure 4.** (A) Reaction scheme of the fluorinase catalysed transhalogenation of **26** to [ $^{18}\text{F}$ ]FDA-PEG-biotin ([ $^{18}\text{F}$ ]**27**). (B) HPLC radioactivity trace of a fluorinase catalysed conversion of CIDA-PEG-biotin **26** to [ $^{18}\text{F}$ ]**27** in the presence of  $^{18}\text{F}^-$ , L-Se-Met in phosphate buffer (pH 7.8), at 37 °C after 30 min incubation. [ $^{18}\text{F}$ ]**27** was observed at  $t_R = 9.7$  min, showing 40% radiochemical conversion during this experiment.

After a 30 min incubation,<sup>[15]</sup> the enzyme was cleanly precipitated by heat denaturation and the solution removed after centrifugation, prior to purification by HPLC. During semi-prep HPLC the eluted peak corresponding to [ $^{18}\text{F}$ ]**27** was collected (Figure 4,  $t_R = 9.7$  min), diluted with water (50 mL) and loaded onto a  $\text{C}_{18}$  reverse phase cartridge, which was then washed with water (20 mL) and [ $^{18}\text{F}$ ]**27** eluted with ethanol. The purity of [ $^{18}\text{F}$ ]**27** was determined by analytical radio-HPLC. This revealed the existence of an additional radioactive peak at  $t_R = 4.3$  min (Figure 5). A typical procedure from [ $^{18}\text{F}$ ]fluoride (457 MBq) to the elution of [ $^{18}\text{F}$ ]**26** (59.7 MBq) took 1 h 19 min. After semi-prep HPLC and  $\text{C}_{18}$  cartridge purification, [ $^{18}\text{F}$ ]**27** was recovered with a radiochemical yield of 13% (decay uncorrected) and a radiochemical purity of 90%.



**Figure 5.** Analytical HPLC radio trace of  $[^{18}\text{F}]\text{FDA-PEG-biotin}$  ( $[^{18}\text{F}]\mathbf{27}$ ) acquired using standard purification protocols (red) and optimized purification protocols (black). Structures of  $[^{18}\text{F}]\text{FDA-PEG-biotin}$  ( $[^{18}\text{F}]\mathbf{27}$ ,  $t_R = 5.7$  min) and  $[^{18}\text{F}]\text{FDA-PEG-biotin sulfoxide}$  ( $[^{18}\text{F}]\mathbf{28}$ ,  $t_R = 4.3$  min) are shown. For full conditions see the Experimental Section and Supporting Information.

### Optimisation of $[^{18}\text{F}]\text{FDA-PEG-biotin}$ purification protocol

The chemical identity of the impurity (10%,  $t_R = 4.3$  min, Figure 5) in the analytical radio-HPLC trace was elucidated by mass spectrometry analysis. An equivalent peak could be observed in nonradioactive storage of a solution of **26** in MeCN/H<sub>2</sub>O at room temperature where the conversion to product **28** is slow (3% at 48 h). Isolation by semi-prep HPLC and analysis by LC-MS found a  $[\text{M}+\text{H}]^+$  mass of 741 amu, consistent with FDA-PEG-biotin sulfoxide (**28**) (see Figure 5). Indeed, previous studies concerning the radiolabelling of biotin observed a similar outcome.<sup>[21]</sup> Furthermore, biotin sulfoxide is a known product in mammalian catabolism, and is observed in both urine and plasma.<sup>[22]</sup> The sulfoxide is also known to bind avidin,<sup>[23]</sup> and thus ( $[^{18}\text{F}]\mathbf{28}$ ) low levels are unlikely to negatively affect PET studies, however we sought to exclude it from the biotransformation. Studies focused on the radiolytic decomposition of concentrated radiopharmaceuticals<sup>[24]</sup> indicate that reactive oxygen species (hydroxyl radicals and/or hydrogen peroxide) can be detrimental, and such species could clearly account for the sulfur oxidation of  $[^{18}\text{F}]\mathbf{28}$ . In an effort to minimize the formation of  $[^{18}\text{F}]\mathbf{28}$ , the fluorinase mediated radiolabelling was repeated however in this instance making use of degassed eluting solvents and established anti-oxidant stabilisers. Approximately the same levels of radioactivity ( $\sim 70$  MBq) was collected by semi-prep HPLC into a vial containing 10 mg/mL ascorbate (a known radical scavenger which is amenable to human use) in water solution (45 mL, degassed). This solution was loaded directly onto the C18 cartridge which had been pre-equilibrated with the same degassed solution and  $[^{18}\text{F}]\mathbf{27}$  was then eluted with ethanol (0.4 mL). HPLC analysis of the  $[^{18}\text{F}]\mathbf{27}$  revealed it to be free of  $[^{18}\text{F}]\mathbf{28}$ , and with a radiochemical purity of  $>99\%$  (see Figure 5). Furthermore, the stability of  $[^{18}\text{F}]\mathbf{27}$  in the ethanol fraction (left open to the atmosphere) was monitored over time by using formulations of both water and aqueous ascorbate (10 mg/mL) solution. This did not reveal any appreciable oxidation of  $[^{18}\text{F}]\mathbf{27}$ . Under these optimised conditions there was no observable degradation of either  $[^{18}\text{F}]\mathbf{27}$  or its chlorinated precursor. Such stability affords the prospect of the

development of an automated recycling protocol, potentially incorporating immobilised enzyme (to circumvent the need for centrifugation), to be used in the synthesis of [<sup>18</sup>F]**27** to allow for the formulation of higher doses. This fluorinase mediated <sup>18</sup>F-labelling methodology provides a novel route into [<sup>18</sup>F]biotin in the form of [<sup>18</sup>F]**27**, and demonstrates a direct 'last step' labelling approach through the formation of a chemically stable C-<sup>18</sup>F bond from a chlorinated precursor. Furthermore, after optimization of the purification protocol with the use of non-toxic radical scavengers, a radiochemically pure sample of [<sup>18</sup>F]**27** was achieved.

### **[<sup>18</sup>F]FDA-PEG-biotin streptavidin binding experiments**

To confirm the utility of [<sup>18</sup>F]**27** as a candidate PET tracer for clinical use an assessment of its streptavidin binding affinity was performed. Accordingly [<sup>18</sup>F]**27** was incubated in PBS buffer (pH 7.8) in the presence and absence of streptavidin (see Supporting Information). Thin layer chromatography was then conducted on both samples (+/- streptavidin) which were analysed using a radio-TLC scanner to determine the percentage of streptavidin bound [<sup>18</sup>F]**27**. The radio-traces obtained from samples [<sup>18</sup>F]**27** incubated without streptavidin revealed a broad peak with a R/F value of 0.39. Upon incubation with streptavidin [<sup>18</sup>F]**27** is almost completely retained (above 96%) at the origin of the TLC plate (R/F value of 0.02, see Supporting Information) indicating its almost complete complexation with streptavidin. This finding is in accordance with the high affinity binding of non radioactive FDA-PEG-Biotin **27** and avidin.

## **Conclusions**

A FDA-PEG-tetrazine construct **17** and a radiolabelled (+)-biotin derivative [<sup>18</sup>F]**27** were generated from their CIDA-chlorinated precursors **16** and **26** respectively. In both cases each was shown to efficiently couple to models of their complementary partners anticipated for *in vivo* studies. As part of the study a protocol to avoid radiolytically triggered oxidation of [<sup>18</sup>F]**27** was developed. This adds to the repertoire of conjugates for use in fluorinase catalysed radio-synthesis for PET and shows that the fluorinase will accept a wide range of CIDA substrates tethered at C-2 of the adenine ring with a pegylated cargo.

## **Experimental Section**

### **Cold Transhalogenation assay**

A typical cold transhalogenation assay is exemplified in **26**. In a total reaction volume of 1000  $\mu$ L (in 50 mM phosphate buffer, at pH 7.8), recombinant fluorinase (0.8 mg/mL<sup>-1</sup>) was incubated with CIDA-PEG-biotin **26** (0.1 mM), L-Se-Met (0.075 mM) and KF (50 mM) at 37 °C. Samples (50  $\mu$ L) were periodically removed, the protein precipitated by heating at 95 °C for 5 min, before being clarified by centrifugation (13 000 rpm, 10 min). Samples of the supernatant (40  $\mu$ L) were removed for analysis by HPLC. HPLC analysis was performed on a Shimadzu Prominence system using a Kinetix 5  $\mu$ m XB-C18

100A (150 mm × 4.6 mm) column and a guard cartridge. Mobile phase: 0.05% TFA in water (solvent A) and 0.05% TFA in MeCN (solvent B); Linear Gradient: 15% solvent B to 95% solvent B over 25 min, 95% for 5 min, and back to 15% B for 10 min to re-equilibrate the column. Flow rate: 1 mL/min<sup>-1</sup>; Detection: 254 nm; Injection volume: 40 μL.

### HABA Assay

Preliminary experiments were performed varying the concentrations of HABA and avidin to determine the optimal conditions for the competitive binding studies. In the final experiments a range of concentrations (0-200 μM) of FDA-PEG-Biotin **27** and (+)-biotin were added to a HABA/avidin complex, obtained by mixing avidin and HABA to final concentrations of (2.5 mg/ml) and 0.25 μM respectively. Changes in absorbance were measured at 500 nm and the % inhibition was calculated using the following formula:  $\frac{[(\text{absorbance with competitor}) - (\text{absorbance without HABA})]/(\text{Av complex})}{[(\text{absorbance without competitor}) - (\text{absorbance without HABA})]/(\text{Av complex})} \times 100$ .

### [<sup>18</sup>F] Labelling of **26** to [<sup>18</sup>F]**27**

A typical <sup>18</sup>F labelling experiment of **26** was performed as follows: L-selenomethionine (40 μL of a 2mM solution in water) and compound **26** (0.4 mg in 50 μL of water) were added successively to an Eppendorf tube containing a solution of fluorinase (5 mg in 50 mm phosphate buffer, 110 μL). The contents were mixed well with a pipette and to this mixture was added [<sup>18</sup>F]fluoride in [<sup>18</sup>O]water (457 MBq, 50 μL), making a total volume of 250 μL. The contents were again well mixed and incubated at 37 °C for 30 min. After this time the reaction was stopped and the mixture denatured by heating at 95 °C for 5 min and water (400 μL) added before being clarified by centrifugation (13 000 rpm, corresponding to 16060 g, 5 min). The supernatant was injected into a Shimadzu Prominence HPLC system equipped with a quaternary pump, a degasser, a flow cell detector and a diode array detector using a Phenomenex Kingsorb C18 (250 × 10.00 mm, 5 μm) column and a guard cartridge. Mobile phase: 0.05% TFA in water (solvent A) and 0.05% TFA in MeCN (solvent B); Linear Gradient: 15% solvent B to 38% solvent B over 16 min, 95% for 5 min, and back to 15% B for 10 min to re-equilibrate the column. Flow rate: 5 mL/min<sup>-1</sup>. The radioactive fraction corresponding to the reference of [<sup>18</sup>F]**27** was collected, diluted with 10 mg/mL ascorbate in water (50 mL) and loaded onto a preactivated Waters Oasis HLB® Cartridge (conditioned with 2 mL EtOH and 5 mL 10 mg/mL ascorbate/water). The cartridge was washed with 20 mL of 10 mg/mL ascorbate/water and the desired product was collected by eluting with 0.6 mL of ethanol, to give 59.7 MBq (13 %, decay uncorrected) of >99% pure product of [<sup>18</sup>F]**27**.

### Streptavidin binding Radio TLC assay

To PBS buffer (300 μL, pH 7.8) or streptavidin (0.6 mg) in PBS buffer (300 μL, pH 7.8) was added [<sup>18</sup>F]**27** (0.8 MBq, in a 50 μL solution of 1:5 EtOH:water), the reaction mixture was mixed well and incubated at 37 °C for 30 min. Thin layer chromatography was then conducted on both samples (+/- streptavidin) using aluminium plates coated with silica gel (60F<sub>245</sub> Merck), eluted with a solution 75:25:5

MeCN:MeOH:Water. TLC plates were examined using a radio-TLC scanner to determine the percentage of streptavidin bound [<sup>18</sup>F]**27**. Thin layer chromatograms radio-traces of the samples [<sup>18</sup>F]**27** in PBS buffer revealed an R/F value of 0.39, upon incubation with streptavidin thin layer chromatograms radio-traces indicated almost completely retention of [<sup>18</sup>F]**27** (about 96%) in the origin of the TLC plate (R/F value of 0.02, see Supporting Information for full experimental details and Figures).

**See Supporting Information for experimental detail on:** Compound synthesis and characterisation, assay conditions and fluorinase overexpression and purification.

## Acknowledgements

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**Keywords:** fluorinase · amide bioconjugation · tetrazine · biotin · <sup>18</sup>F labelling

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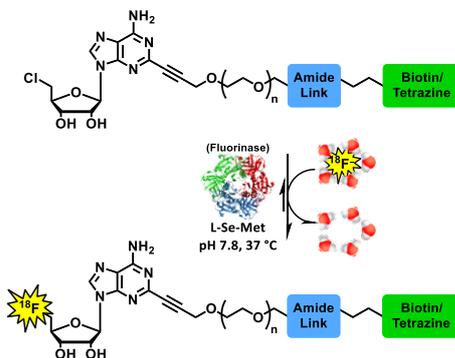
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## Entry for the Table of Contents

### FULL PAPER

The synthesis and evaluation of new amine and acid functionalized fluorinase substrates, allowing for the attachment to an extensive range of carboxylate or amine containing bioactive molecules to a fluorinase binding 5'-CIDA moiety, prior to their enzymatic radiolabelling with [ $^{18}\text{F}$ ]fluoride. Two new substrates of this class have been designed specifically for their application towards a pretargeting approach for radiolabelling conjugated antibodies and proteins for PET imaging.



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**Enzymatic fluorination of biotin and tetrazine conjugates for pre-targeting approaches to PET imaging.**