DOI: 10.1111/1471-0528.17386

SYSTEMATIC REVIEW



DG An International Journal of Obstetrics and Gynaecology

The accuracy of cell-free DNA screening for fetal segmental copy number variants: A systematic review and meta-analysis

Yvette C. Raymond¹ | Melissa L. Acreman² | Sofia Bussolaro³ | Ben W. Mol^{1,4}

Ilaria Fantasia¹⁰ Daniel Lorber Rolnik^{1,5}

Shavi Fernando^{1,5} Melody Menezes^{6,7} Fabricio Da Silva Costa^{8,9}

¹Department of Obstetrics and Gynaecology, Monash University, Clayton, Victoria, Australia

²Department of Obstetrics and Gynaecology, Ipswich Hospital, Ipswich, Queensland, Australia

³Department of Medicine, Surgery and Health Sciences, University of Trieste, Trieste, Italy

⁴Aberdeen Centre for Women's Health Research, University of Aberdeen, Aberdeen, UK

⁵Monash Women's, Monash Health, Clayton, Victoria, Australia

⁶Monash Ultrasound for Women, Melbourne, Victoria, Australia

⁷Department of Paediatrics, The University of Melbourne, Melbourne, Victoria, Australia

⁸Maternal Fetal Medicine Unit, Gold Coast University Hospital, Gold Coast, Queensland, Australia

9School of Medicine, Griffith University, Gold Coast, Queensland, Australia

¹⁰Obstetrics & Gynaecology Unit, San Salvatore Hospital, L'Aquila, Italy

Correspondence

Yvette C. Raymond, 6/47 Carroll Crescent, Glen Iris, Vic. 3146, Australia. Email: yvette.raymond@monash.edu

Abstract

Background: The performance of cell-free DNA (cfDNA) screening for microscopic copy number variants (CNVs) is unclear.

Objectives: This was a systematic review and meta-analysis to investigate the sensitivity, specificity and positive predictive value (PPV) of cfDNA screening for CNVs.

Search Strategy: Articles published in EMBASE, PubMed or Web of Science before November 2022 were screened for inclusion. This protocol was registered with PROSPERO (23 March 2021, CRD42021250849) prior to initiation.

Selection Criteria: Articles published in English, detailing diagnostic outcomes for at least 10 high-risk CNV results with cfDNA were considered for inclusion.

Data Collection and Analysis: The PPV was calculated and pooled with randomeffects models for double-arcsine transformed proportions, using cases with diagnostic confirmation. Overall sensitivity, specificity and a summary receiveroperating characteristics (ROC) curve were calculated using bivariate models. The risk of bias was assessed using QUADAS-2.

Main Results: In all, 63 articles were included in the final analysis, detailing 1 591 459 cfDNA results. The pooled PPV was 37.5% (95% confidence interval [CI] 30.6-44.8), with substantial statistical heterogeneity ($I^2 = 93.9\%$). Bivariate meta-analysis estimated sensitivity and specificity to be 77.4% (95% CI 65.7-86.0) and 99.4% (95% CI 98.0-99.8), respectively, with an area under the summary ROC curve of 0.947 (95% CI 0.776-0.984).

Conclusions: Approximately one-third of women who screen high-risk for CNVs with cfDNA will have an affected fetus. This value is of importance for screening counselling.

KEYWORDS cell-free DNA, cell-free DNA screening, segmental copy number variants

INTRODUCTION 1

Cell-free DNA (cfDNA) screening is currently the most accurate form of prenatal screening for fetal aneuploidy. Compared with other screening methods, cfDNA screening (also termed non-invasive prenatal testing [NIPT]) has demonstrated an impressively high sensitivity in detecting common fetal genetic anomalies such as trisomy 21, alongside a lower false-positive rate.^{1,2} Importantly, however, cfDNA screening may still return incorrect results, and thus

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2023 The Authors. BJOG: An International Journal of Obstetrics and Gynaecology published by John Wiley & Sons Ltd.

is not suitable as a diagnostic test.^{3,4} More recently, cfDNA screening panels have been expanded beyond common aneuploidies to include segmental copy number variants (CNVs), which encompass sub-microscopic deletions and duplications.

The phenotypical consequences of CNVs vary significantly, depending on size of the variant and the gene region involved, but range from completely benign to incompatible with life.^{5,6} Several clinical syndromes have been identified as being attributable to specific chromosomal microdeletions, including 22q.11.2 syndrome (previously termed DiGeorge syndrome), 15q.11 microdeletion (Angelman) syndrome, and 5p- (Cri Du Chat) syndrome.^{7–9}

Individual CNVs, such as 22q.11.2 syndrome, are generally rare.⁷ When considered as a collective, their frequency increases up to 6% in fetuses with anatomical anomalies identified on ultrasound.¹⁰ They also occur unpredictably, with 90–95% of 22q.11.2 syndrome diagnoses attributable to de novo aberrations, which are more likely to be pathological.^{7,11} This, in combination with the absence of traditional identifiable risk factors for fetal genetic anomalies, such as advanced maternal age, which has no correlation with CNVs, makes a reliable prenatal screening method for CNVs desirable.^{12,13}

To date, the performance of cfDNA screening for CNVs has been less than ideal. Several studies have documented a significantly lower positive predictive value (PPV) for CNVs compared with common aneuploidies.^{14–18} Similarly, the sensitivity of cfDNA screening for CNVs appears suboptimal.⁶ There is, however, little consensus regarding these estimates, with values for both PPV and sensitivity varying dramatically across studies.¹⁹ This systematic review and meta-analysis aims to investigate the diagnostic accuracy of cell-free DNA screening for CNVs.

2 | METHODS

We conducted a systematic review of the literature and metaanalysis of diagnostic accuracy to assess cfDNA screening for fetal microscopic CNVs in the general obstetric population, using results from prenatal or postnatal cytogenetic diagnostic tests as validation. The protocol for this review was registered with the International Prospective Register of Systematic Reviews (PROSPERO) (23 March 2021, CRD42021250849), prior to its initiation, and results are reported in accordance with the Preferred Reporting Items for Systematic reviews and Meta-Analyses (PRISMA) statement.²⁰

2.1 | Eligibility criteria

Studies eligible for inclusion were original research articles or abstracts reporting the performance of cfDNA screening for fetal CNVs in a pregnant population. Studies that reported fetal diagnostic confirmation for <10 high-risk results were excluded.^{21,22} Animal studies, systematic reviews, metaanalyses, case-reports and articles not published in English (except where English translation was available) were excluded.

2.2 | Information sources, search strategy and selection process

PubMed, EMBASE and Web of Science were searched from inception to November 2022. The full search strategy, which was designed to investigate cfDNA screening for conditions other than the common trisomies as part of a broader research project, is provided in Appendix S1. Study selection was conducted using Covidence systematic review software (Veritas Health Innovation). Each title and abstract were independently reviewed for inclusion by two investigators (MA, SB or YR), with a third investigator consulted in case of disagreement (DR, FC or IF). Full texts of potentially eligible articles were then reviewed in a similar process. References of included articles were also manually screened to identify articles potentially missed in the search.

2.3 Data collection process and data items

Following full-text review, data were manually extracted by one investigator per article (MA, SB or YR). In instances in which desired data were missing, investigators attempted to contact corresponding authors via email on two subsequent occasions, after which articles were excluded if no response was obtained. Information extracted included author names, year of publication, publication title, country, study design, anomalies screened for, populations screened, proportion of high-risk cfDNA screening results with and without diagnostic follow-up, and reported screening accuracy.

Studies that reported pregnancy outcomes only for highrisk cfDNA results were considered case series; those that reported the performance of cfDNA screening using case and control groups of fetuses with previously determined karyotypes were classified as case-control studies, and studies that prospectively assessed pregnancy outcome for both high- and low-risk cfDNA screening results were classified as cohort studies.

The populations screened in each article were categorised as predominately low-risk (<50% of individuals with high baseline risk of fetal CNV) or high-risk (\geq 50% of individuals with high baseline risk of fetal CNV). This delineation was made, as the majority of studies only provided grouped statistics for the population screened, thus we were unable to assess participant background risk as a continuous variable. Recognised risk factors for CNVs included high-risk serum or combined screening results, fetal anomalies on ultrasound examination, and prior history of chromosomal anomalies. Advanced maternal age was not considered a risk factor.²³

2.4 | Risk of bias assessment

Risk of bias assessment for each included study was conducted independently by two investigators (MA, SB or YR), with discordant results resolved by consensus, using the Quality Assessment of Diagnostic Accuracy Studies 2 (QUADAS-2) tool.²⁴ For the 'patient selection' domains, the aforementioned categorisation of high- versus low-baseline aneuploidy risk was used to determine bias and applicability risks. Additionally, any studies in which participant selection was contrived (not random or sequential) were deemed high-risk of bias.

In the 'index test' domain, risk of bias was classified as high when fetal karyotype was known prior to cfDNA screening. Studies in which the cfDNA screening methodology or platform was not specified were labelled unclear for both bias and applicability.

For the 'reference standard' domains, studies were deemed high-risk for bias and applicability if >10% of diagnostic confirmations were ascertained by chorionic villus sampling (CVS), as these could include confined placental mosaicism and our analysis pertains to fetal outcomes. Additionally, studies in which >10% of diagnostic investigations were conducted by karyotyping (as opposed to microarray or high-depth sequencing), which may not detect sub-microscopic anomalies or in which the cytogenetic methods were unspecified, were considered high- and unclear risk of bias, respectively.

For the 'flow and timing' domain, the threshold for low risk of bias was arbitrarily set at \geq 80% diagnostic follow-up rate.

2.5 | Effect measures, synthesis methods and statistical analysis

Positive predictive values were calculated using results from case series and cohort studies and excluding case-control studies. PPV were calculated as the number of true-positive cfDNA results validated by a diagnostic investigation, over the total number of results with diagnostic follow-up. Screening results with no genetic confirmation were excluded from the analysis. The PPV was calculated for CNVs overall, and individually for the most common CNV syndromes including 22q.11.2, 15q.11 microdeletion and 5p.

To achieve stabilisation of the variances and because some studies had PPV of 0% or 100%, estimates were transformed using the Freeman–Tukey double-arcsine transformation. The transformed proportions were then pooled with random-effects models using inverse-variance weights and the DerSimonian and Laird method to estimate the betweenstudy heterogeneity.

We assessed heterogeneity by calculating the I^2 statistic. To explore heterogeneity, univariable mixed-effects metaregression models were fit to the data using predictors including year of publication, diagnostic test follow-up rates and baseline population risk category. Random-effects meta-analyses of subgroups according to baseline population risk (\geq 50% or <50% high-risk) and diagnostic follow-up rate (\geq 80% or <80%) were also conducted to investigate subgroup effects. With the aim of investigating the impact of bias on PPV estimates, sensitivity analysis was performed including only studies considered to be at low risk in all four 'bias' domains of the QUADAS-2 tool.

Sensitivity and specificity were calculated using results from case-control studies and cohort studies with diagnostic confirmation ascertained for both high- and low-risk screening results. A bivariate random-effects model was then used to estimate pooled sensitivity and specificity and create a summary receiver-operating characteristics (sROC) curve. The 95% confidence interval (95% CI) for the area under the sROC curve was estimated using 5000 bootstrap samples.

Publication bias and small-study effects were investigated through inspection of a funnel plot and with Egger's test. Analyses were conducted with the packages 'metafor' and 'mada' in R, and *p*-values < 0.05 were considered statistically significant.²⁵

3 RESULTS

3.1 Study characteristics

In total, 7845 search results were identified, of which 1862 were duplicates. After screening the remaining 5983 results, 63 articles satisfied the inclusion criteria.^{12,14–16,18,26–83} The study selection process is shown in Figure 1. Authors were contacted for seven articles with incomplete data; data were obtained for one³⁸ and six were excluded.^{84–89}

Included studies were published between 2015 and 2022. Across all included studies, 1 591459 women underwent cfDNA screening for CNVs, with 5481 receiving a high-risk result (screen-positive rate of 0.34%). This screen-positive rate may be over or underestimated, however, as some studies (n = 9) did not report the total number of women screened (thus reducing the denominator), or the number of women who received a high-risk result without diagnostics (n = 8) (thereby reducing the numerator). Diagnostic results were available for 3737 pregnancies that screened high-risk for a CNV (68.2%), including 934 at high-risk for one of the deletion syndromes (22q.11.2 syndrome, n = 632; 15q microdeletion, n = 179; 5p- syndrome, n = 123).

Of the 63 included studies, PPV results were calculated from 59 (48 case-series,^{12,14–16,18,26–28,30,33–38,40–46,48,49,52–59, ^{61–66,68–71,74,78–82} and 11 cohort studies^{29,47,50,60,67,72,73,75–77,83}). Sensitivity and specificity were reported from nine (four case-controls,^{31,32,39,51} five cohort studies with complete diagnostic confirmation^{29,60,75–77}). Twenty-five of the 63 included studies (39.7%) involved predominately high-risk cohorts.^{15,26,28,29,31,35,37,39,41,42,45,51,54,56,57,59,60,62,65,67,71,73–75,78} Six inclusions (9.5%) were validation studies assessing novel cfDNA technologies or algorithms.^{15,29,31,39,47,49}}

Seventeen studies (27.0%) reported cfDNA screening outcomes for 22q.11.2 syndrome (16 reporting $PPV^{12,14,26,27,30,38,40,47,56,60,65,68,69,73,76,83}$ and three reporting sensitivity and specificity^{39,60,76}), including seven which

551

552



FIGURE 1 PRISMA flowchart of study inclusion. CNV, copy number variant; RAT, rare autosomal trisomy, SCA, sex chromosome aneuploidy.

exclusively assessed 22q.11.2 syndrome and no other CNVs.^{27,30,39,60,68,69,76} Six studies (9.5%) included results for 15q microdeletion,^{12,14,26,38,40,73} and eight (12.7%) for 5p-syndrome.^{12,14,26,38,40,47,73} A summary of the characteristics of included studies is provided in Table S1.

3.2 | Risk of bias

The outcomes of the bias assessment using the QUADAS-2 tool are depicted in Figure 2 and Figure S1. The most

frequent high-risk bias arose in the 'Flow and Timing' domain as a result of incomplete diagnostic confirmation of cfDNA results, with 31 studies (49.2%) deemed high-risk. ^{15,18,26–28,30,33–35,37,38,42,45–49,52,54,55,58,62–65,70,71,73,74,82,83} This is most likely attributable to loss of follow-up of study participants or maternal decline of diagnostic testing. The most frequent applicability concern was in 'Reference Standard', with 15.9% (n = 10/63) of studies being classified as high-risk. ^{14,31,33,36,39,51,54,56,60,73} Only six studies (9.5%) were deemed to be low-risk in all four of the QUADAS-2 bias domains. ^{50,53,68,72,78,79}





FIGURE 2 Pooled domain outcomes of the QUADAS-2 bias assessment tool.

Examination of the funnel plot (Figure S2) revealed no evidence of publication bias, which was consistent with the Egger's test results (p = 0.30).

3.3 | Positive predictive value

The PPV of cfDNA screening for CNVs was generated using results from 3715 women with diagnostic confirmation. Meta-analysis revealed a pooled PPV of 37.5% (95% CI 30.6–44.8). The forest plot for PPV is shown in Figure 3. On sensitivity analysis for PPV using only results from the six studies deemed low risk in all four bias domains, the PPV was 34.3% (95% CI 22.3–47.4; Figure S3).

There was substantial statistical heterogeneity, with an I^2 statistic of 93.9% (p < 0.01). Meta-regressions were conducted for baseline cohort risk, year of publication and diagnostic follow-up rate; however, none significantly explained heterogeneity (p = 0.477, 0.445 and 0.327, respectively).

Subgroup analyses with baseline cohort risk and proportion of diagnostic confirmation revealed that the PPV in predominately low-risk cohorts (n = 53/59) was 36.0% (95% CI 28.9–43.3), and 51.5% (95% CI 24.9–77.8) in predominately high-risk cohorts (n = 6/59, p-value for subgroup difference 0.29). In studies with ≥80% proportion of diagnostic follow-up (n = 28/59), the PPV was 33.8% (95% CI 24.8–43.2), and for studies with diagnostic follow-up <80%, the PPV was 41.0% (95% CI 30.7 –51.8) (n = 31/59, p-value for subgroup difference 0.32). The forest plots for subgroup analyses are provided in Figures S4 and S5.

For 22q.11.2 syndrome, subtype meta-analyses of the 16 studies (n = 1/16 with predominately high-risk cohorts, 8/16 with <80% diagnostic follow-up) detailing 622 women with diagnostic confirmation, revealed a PPV of 49.0% (95% CI 24.5–73.7) (Figure S6). The PPV for 15q microdeletion syndrome based on six studies (n = 0/6 with predominately high-risk cohorts, 3/6 with <80% diagnostic follow-up) and 179 women was 25.9% (95% CI 0.0–76.6) (Figure S7). The PPV for 5p- based on the results of eight studies (n = 0/8 with predominately high-risk cohorts, 5/8 with <80% diagnostic follow-up) and 123 women was 30.8% (95% CI 9.1–56.8) (Figure S8). The heterogeneity amongst these three subtype analyses was high, with I^2 values of 96.2%, 92.8% and 76.4% for 22q.11.2 syndrome, 15q.11 microdeletion syndrome and 5p- syndrome, respectively.

3.4 | Sensitivity and specificity

Bivariate meta-analysis of sensitivity and specificity consisted of nine studies (four case-controls with 118 cases and 2551 controls,^{31,32,39,51} five cohort studies with 32 322 women^{29,60,75-77}), including six with predominately high-risk cohorts.^{29,31,39,51,60,75} Individual estimates from these studies are presented in Figure S9. Bivariate meta-analysis estimated overall sensitivity to be 77.4% (95% CI 65.7–86.0) and specificity to be 99.4% (95% CI 98.0–99.8). These values generated an area of 0.947 (95% CI 0.776–0.984) under the summary ROC curve, shown in Figure 4.

The sensitivity and specificity of 22q.11.2 syndrome were not meta-analysed due to these outcomes only being reported by three articles; however, reports ranged between 69.6% and 90.0% for sensitivity, and 99.7% and 100.0% for specificity.^{39,60,76}

4 | DISCUSSION

4.1 | Main findings

This meta-analysis revealed that the overall PPV of cfDNA screening for fetal CNVs is <40%. The probability of fetal confirmation is slightly lower among women with a low baseline risk of aneuploidy (32%) when estimates are generated using only results from studies with high rates of diagnostic confirmation (34%), and at low risk of bias (32%), although these differences were not statistically significant. The sensitivity of cfDNA screening for CNVs is 77%, and the specificity is over 99%. Another major finding of this review is the substantial heterogeneity among included studies.

4.2 Interpretation

It is probable that the pooled PPV of 38% observed in this meta-analysis is overestimated by biases amongst included studies. We observed a reduction in PPV when only studies at low risk of bias were considered, and although this reduction was not significantly lower than the pooled PPV estimate, it does suggest that well-conducted studies lead to lower PPV estimates. Two prominent bias sources likely causing overestimation of the PPV include baseline cohort risk of CNVs and diagnostic confirmation rate.





1341

3715

0

20

40

Positive Predictive Value (%)

60

80

100

Pre-screening probability of fetal CNV is higher among women with risk factors, which increases the PPV.90 Similarly, women with higher suspicion of CNVs are more

 $I^2 = 94\%, \, \tau^2 = 0.0675, \, \rho < 0.01$

Overall

likely to undergo diagnostic testing, thus incomplete diagnostic follow-up may lead to a greater proportion of high-risk pregnancies in the follow-up cohort.⁹¹

37.54 [30.59; 44.75] 100.0%

1.7%

1.4%

1.5%

1.9%

1.8%

1.6%

1.5%

1.4%

1.8%

1.4%

1.6%

1.9%

17%

1.9%

1.5%

1.8%

1.6%

1.6%

1.5%

1.7%

1.9%

1.6%

1.7%

1.6%

1.7%

1.9%

14%

1.9%

1.5%

1.7%

1.8%

1.7%

1.7%

1.6%

1.5%

1.9%

1.8%

1.9%

1.8%

1.6%

1.8%

1.8%

1.6%

1.5%

1.6%

1.8%

1.9%

1.8% 1.8%

1.9%

1.7%

1.5%

1.5%

1.8%

1.8%

1.8%

1.9%

1.9%

1.7%



FIGURE 4 Summary receiver-operating characteristics (ROC) curve of cell-free DNA screening in the detection of copy number variants. The grey triangles represent estimates from nine individual studies, the closed circle represents the pooled estimate, and the dotted ellipse represents the 95% confidence region.

None of the tested variables explained the substantial heterogeneity among studies. For baseline cohort risk, this may be due to our categorisation of 'high risk' versus 'low risk' using an arbitrary cut-off value of 50%. For diagnostic confirmation rates, heterogeneity may arise between studies with similar follow-up depending on the diagnostic methods utilised, as some cytogenetic investigations such as karyotype are less reliable in detecting CNVs, and CVS may provide a false indication of fetal involvement, particularly when the ultrasound is normal. There are also several other factors that may contribute to heterogeneity, including cfDNA screening platform, technology, depth of sequencing and range of sizes of detected anomalies.

Results for individual CNV conditions were markedly more uncertain than those for grouped CNVs, with wider confidence intervals. The pooled PPV for 22q.11.2 syndrome was also markedly higher than that of grouped CNVs. This is likely attributable to the small cohort sizes, in which even a small number of true-positive results have the power to inflate PPV. There was also significant potential for bias among these studies, with only one study investigating 22q.11.2 syndrome at low risk of bias, and no studies investigating either 5p- or 15q.11 microdeletion syndromes at low risk of bias.

The sensitivity observed is considerably lower than that of cfDNA screening for common trisomies, as almost onequarter of CNVs may go undetected by cfDNA screening, based on our findings.¹ This is likely attributable to smaller aberration sizes of many CNVs compared with whole chromosome aneuploidies, in tandem with the cost and technology restraints for sequencing depth in commercial cfDNA screening. There was one predominate outlier which reported a sensitivity of 46.1%, although this study consisted exclusively of high-risk results for microdeletion syndromes other than 22q.11.2.⁷⁷ The specificity revealed in this metaanalysis was high, despite the presence of one outlier which reported a specificity of 87.5%.⁷⁵

U

Obstetrics and Gyr

4.3 Comparison with previous studies

To the best of our knowledge, this is the first meta-analysis investigating the performance of cfDNA screening for CNVs. A previous systematic review was conducted by Familiari et al. in which cfDNA screening was investigated exclusively for microdeletions or microduplications, and notably only in large cohorts with >5000 women. Despite a smaller number of included studies due to different search methodologies, researchers found a similar PPV as that for CNVs in this study, of approximately 40%, based on the results of seven studies.¹⁹ Similar to the results of our meta-analysis, this study was also plagued by substantial heterogeneity, with PPV ranging between 29% and 91%.

4.4 | Clinical implications

A reliable screening method for CNVs is desirable, particularly for the CNV syndromes, as these conditions have profound impacts on health and development. Nevertheless, the results of this meta-analysis demonstrate that the clinical implementation of extended cfDNA screening panels should be approached with caution.

Based on the results of this analysis, approximately onethird of women who receive a high-risk result will have an affected fetus. It should, however, be stressed that this is likely an overestimate, given the high degree of heterogeneity and biases affecting included studies. While low PPV is expected in screening for rare diseases despite reasonable sensitivity and high specificity, the clinical implications of false-positive results are not negligible. These include significant parental anxiety and procedural-related risks of diagnostic investigations which should not be overlooked.² Even when fetal anomalies are observed on ultrasound prior to screening, diagnostic testing is arguably a more appropriate investigation for these pregnancies, as cfDNA is only appropriate for screening.

While the PPV is considerably lower than that of common aneuploidies such as trisomy 21, expanded screening for CNVs is defensible in the event of successful identification of a clinically significant anomaly that would otherwise be missed prenatally. However, the sensitivity revealed in this meta-analysis suggests that almost one-quarter of CNVs may be undetected by screening. Additionally, the benefit of screening for these anomalies is often questionable even in the event of successful identification, as the clinical consequences of many CNVs are poorly understood.^{5,6} This creates a challenge for clinicians to provide genetic counselling for largely unpredictable phenotypes.

4.5 | Strengths and limitations

The primary strength of this meta-analysis is the number of articles reviewed, with the inclusion of more than 1.5 million women screened. By pooling these results, we were able to obtain estimates of PPV, sensitivity and specificity with relatively high precision. However, in pooling estimates, we are limited by the quality of the included studies which, as demonstrated by our bias assessments, varied considerably.

Another limitation pertains to the arbitrary selection of cut-off values of 50% for high versus low baseline cohort risk, and 80% for diagnostic confirmation rate in the subgroup analyses. While these values were selected to best capture any potential differences between subgroups, it is possible that the division of studies in this way may conceal more subtle gradient effects.

Finally, an integral adjacent to prenatal serum screening in clinical practice is ultrasound examination, as these results have significant influence on pre-test probability and, in turn, PPV. We were limited in this study, as most included articles did not report ultrasound findings, and subsequently we were unable to analyse the association between ultrasound results and screening performance. Similarily, while it is desirable to stratify screening performance by other variables such as the type/size of CNV detected and cfDNA screening technologies utilised, this was not possible, as information on such variables was often not reported.

The performance of cfDNA screening is substantially poorer for CNVs than for common trisomies, with substantial heterogeneity in the literature. Women should be informed about these limitations prior to expanded cfDNA screening, and the low PPV should be carefully considered when counselling women who receive a high-risk result for a CNV.

AUTHOR CONTRIBUTIONS

Conceptualisation: DLR, IF, FDSC. Data curation: YCR, MLA, SB, DLR, IF, FDSC. Formal analysis: DLR, YCR. Investigation: YCR, MLA, SB, DLR, IF, FDSC. Project administration: DLR, IF, FDSC. Supervision: DLR, BM, SF, MM. Visualisation: DLR, YCR. Writing – original draft: YCR. Writing – review & editing: YCR, DLR, IF, FDSC, MLA, SB, BM, SF, MM.

ACKNOWLEDGEMENTS

Open access publishing facilitated by Monash University, as part of the Wiley - Monash University agreement via the Council of Australian University Librarians.

FUNDING INFORMATION

No funding was required or obtained for this research.

CONFLICT OF INTEREST STATEMENT

BWM is supported by a NHMRC Investigator grant (GNT1176437). BWM reports consultancy for ObsEva and Merck and travel support and research grants from

Merck. MM is employed as a genetic counsellor at a private genetic testing provider. DLR has received research grants from NHMRC and Norman-Beischer Medical Research Foundation. The authors declare no competing interests. Completed disclosure of interest forms are available to view online as supporting information.

DATA AVAILABILITY STATEMENT

The datasets and code supporting the current study have not been deposited in a public repository but are available from the corresponding author on request.

ORCID

Yvette C. Raymond D https://orcid.org/0000-0001-8197-7996 Melissa L. Acreman D https://orcid.org/0000-0002-4757-1316 Sofia Bussolaro D https://orcid.org/0000-0002-0273-5700 Ben W. Mol D https://orcid.org/0000-0001-8337-550X Shavi Fernando D https://orcid.org/0000-0001-9160-0084 Melody Menezes D https://orcid.org/0000-0003-3854-0224 Fabricio Da Silva Costa D https://orcid.

org/0000-0002-0765-7780

Ilaria Fantasia ^(b) https://orcid.org/0000-0003-4340-7225 Daniel Lorber Rolnik ^(b) https://orcid.org/0000-0002-2263-3592

REFERENCES

- Gil MM, Accurti V, Santacruz B, Plana MN, Nicolaides KH. Analysis of cell-free DNA in maternal blood in screening for aneuploidies: updated meta-analysis. Ultrasound Obstet Gynecol. 2017;50(3):302–14.
- Akolekar R, Beta J, Picciarelli G, Ogilvie C, D'Antonio F. Procedurerelated risk of miscarriage following amniocentesis and chorionic villus sampling: a systematic review and meta-analysis. Ultrasound Obstet Gynecol. 2015;45(1):16–26.
- Harasim T, Wagner A. Chapter 5 Why cell-free DNA-based noninvasive prenatal testing for fetal chromosome anomalies is not diagnostic. In: Page-Christiaens L, Klein H-G, editors. Noninvasive prenatal testing (NIPT). Cambridge: Academic Press; 2018. p. 67–82.
- Rieder W, White S, McGillivray G, Hui L. Contemporary prenatal aneuploidy screening practice in Australia: frequently asked questions in the cell-free DNA era. Aust N Z J Obstet Gynaecol. 2018;58(4):397–403.
- 5. Zarrei M, MacDonald JR, Merico D, Scherer SW. A copy number variation map of the human genome. Nat Rev Genet. 2015;16(3):172–83.
- 6. Di Renzo GC, Bartha JL, Bilardo CM. Expanding the indications for cell-free DNA in the maternal circulation: clinical considerations and implications. Am J Obstet Gynecol. 2019;220(6):537–42.
- McDonald-McGinn DM, Sullivan KE, Marino B, Philip N, Swillen A, Vorstman JA, et al. 22q11. 2 deletion syndrome. Nat Rev Dis Primers. 2015;1(1):1–19.
- 8. Cerruti Mainardi P. Cri du Chat syndrome. Orphanet J Rare Dis. 2006;1(1):1-9.
- Clayton-Smith J, Laan L. Angelman syndrome: a review of the clinical and genetic aspects. J Med Genet. 2003;40(2):87–95.
- Wapner RJ, Martin CL, Levy B, Ballif BC, Eng CM, Zachary JM, et al. Chromosomal microarray versus karyotyping for prenatal diagnosis. N Engl J Med. 2012;367(23):2175–84.
- Watson CT, Marques-Bonet T, Sharp AJ, Mefford HC. The genetics of microdeletion and microduplication syndromes: an update. Annu Rev Genomics Hum Genet. 2014;15:215–44.
- Petersen AK, Cheung SW, Smith JL, Bi W, Ward PA, Peacock S, et al. Positive predictive value estimates for cell-free noninvasive prenatal screening from data of a large referral genetic diagnostic laboratory. Am J Obstet Gynecol. 2017;217(6):691.e1–6.

- Srebniak MI, Joosten M, Knapen MFCM, Arends LR, Polak M, van Veen S, et al. Frequency of submicroscopic chromosomal aberrations in pregnancies without increased risk for structural chromosomal aberrations: systematic review and meta-analysis. Ultrasound Obstet Gynecol. 2018;51(4):445–52.
- Schwartz S, Kohan M, Pasion R, Papenhausen PR, Platt LD. Clinical experience of laboratory follow-up with noninvasive prenatal testing using cell-free DNA and positive microdeletion results in 349 cases. Prenat Diagn. 2018;38(3):210–8.
- Yang J, Wu J, Peng H, Hou Y, Guo F, Wang D, et al. Performances of NIPT for copy number variations at different sequencing depths using the semiconductor sequencing platform. Hum Genomics. 2021;15(1):41.
- Van Den Bogaert K, Lannoo L, Brison N, Gatinois V, Baetens M, Blaumeiser B, et al. Outcome of publicly funded nationwide first-tier noninvasive prenatal screening. Genet Med. 2021;23(6):1137–42.
- van der Meij KRM, Sistermans EA, Macville MVE, Stevens SJC, Bax CJ, Bekker MN, et al. TRIDENT-2: national implementation of genome-wide non-invasive prenatal testing as a first-tier screening test in the Netherlands. Am J Hum Genet. 2019;105(6):1091–101.
- van Prooyen Schuurman L, Sistermans EA, Van Opstal D, Henneman L, Bekker MN, Bax CJ, et al. Clinical impact of additional findings detected by genome-wide non-invasive prenatal testing: follow-up results of the TRIDENT-2 study. Am J Hum Genet. 2022;109(6):1140–52.
- Familiari A, Boito S, Rembouskos G, Ischia B, Accurti V, Fabietti I, et al. Cell-free DNA analysis of maternal blood in prenatal screening for chromosomal microdeletions and microduplications: a systematic review. Prenat Diagn. 2021;41(10):1324–31.
- Page MJ, McKenzie JE, Bossuyt PM, Boutron I, Hoffmann TC, Mulrow CD, et al. The PRISMA 2020 statement: an updated guideline for reporting systematic reviews. BMJ. 2021;372:n71.
- 21. Abu-Zidan F, Abbas A, Hefny A. Clinical "case series": a concept analysis. Afr Health Sci. 2012;12(4):557–62.
- Patterson PD, Weaver M, Clark S, Yealy DM. Case reports and case series in prehospital emergency care research. Emerg Med J. 2010;27(11):807–9.
- 23. Larroya M, Tortajada M, Mensión E, Pauta M, Rodriguez-Revenga L, Borrell A. Have maternal or paternal ages any impact on the prenatal incidence of genomic copy number variants associated with fetal structural anomalies? PLoS One. 2021;16(7):e0253866.
- Whiting PF, Rutjes AW, Westwood ME, Mallett S, Deeks JJ, Reitsma JB, et al. QUADAS-2: a revised tool for the quality assessment of diagnostic accuracy studies. Ann Intern Med. 2011;155(8):529–36.
- R Core Development Team. R: a language and environment for statistical computing. Vienna: R Foundation for Statistical Computing; 2018.
- Helgeson J, Wardrop J, Boomer T, Almasri E, Paxton W, Saldivar J, et al. Clinical outcome of subchromosomal events detected by whole-genome noninvasive prenatal testing. Prenat Diagn. 2015;35(10):999–1004.
- Saldivar J-S, Monroe T, Boomer T, Wardrop J, Jesiolowski J, Dharajiya N. 334: Identification of 22q11 microdeletions by noninvasive prenatal testing (NIPT) – the clinical experience. Am J Obstet Gynecol. 2015;212(1):S178.
- Taneja P, Curnow K, Halks-Miller M, Seltzer W, de Feo E, Bhatt S. Noninvasive prenatal screening for microdeletions: clinical population and outcomes. Prenat Diagn. 2015;35(Suppl 1):15.
- Yin A-H, Peng C-F, Zhao X, Caughey BA, Yang J-X, Liu J, et al. Noninvasive detection of fetal subchromosomal abnormalities by semiconductor sequencing of maternal plasma DNA. Proc Natl Acad Sci USA. 2015;112(47):14670–5.
- 30. Gross S, Stosic M, McDonald-McGinn D, Bassett A, Norvez A, Dhamankar R, et al. Clinical experience with single-nucleotide polymorphism-based non-invasive prenatal screening for 22q11.2 deletion syndrome. Ultrasound Obstet Gynecol. 2016;47(2):177–83.
- Lefkowitz RB, Tynan JA, Liu T, Wu Y, Mazloom AR, Almasri E, et al. Clinical validation of a noninvasive prenatal test for genomewide detection of fetal copy number variants. Am J Obstet Gynecol. 2016;215(2):227.e1–16.
- 32. Liu H, Gao Y, Hu Z, Lin L, Yin X, Wang J, et al. Performance evaluation of NIPT in detection of chromosomal copy number variants

using low-coverage whole-genome sequencing of plasma DNA. PLoS One. 2016;11(7):e0159233.

- Valderramos SG, Rao RR, Scibetta EW, Silverman NS, Han CS, Platt LD. Cell-free DNA screening in clinical practice: abnormal autosomal aneuploidy and microdeletion results. Am J Obstet Gynecol. 2016;215(5):626.e1–10.
- Ke W, Li Q, Jie S, Chen Q, Zhao W. Non-invasive prenatal DNA testing for genomic copy number variations. Int J Clin Exp Med. 2017;10:5152–9.
- Pescia G, Guex N, Iseli C, Brennan L, Osteras M, Xenarios I, et al. Cell-free DNA testing of an extended range of chromosomal anomalies: clinical experience with 6,388 consecutive cases. Genet Med. 2017;19(2):169–75.
- 36. Williams J, Gupta M, Rad S, Ozimek J, Ratousi D, Farivar S, et al. Positive and negative predictive values of cell-free DNA for noninvasive prenatal testing. Poster abstracts of the ISPD 21st International Conference on Prenatal Diagnosis and Therapy, San Diego, California, USA, 9-12 July 2017. Prenat Diagn. 2017;37(S1):48.
- Liang D, Lin Y, Qiao F, Li H, Wang Y, Zhang J, et al. Perinatal outcomes following cell-free DNA screening in >32 000 women: clinical follow-up data from a single tertiary center. Prenat Diagn. 2018;38(10):755-64.
- Martin K, Iyengar S, Kalyan A, Lan C, Simon A, Stosic M, et al. Clinical experience with a single-nucleotide polymorphism-based non-invasive prenatal test for five clinically significant microdeletions. Clin Genet. 2018;93(2):293–300.
- Ravi H, McNeill G, Goel S, Meltzer SD, Hunkapiller N, Ryan A, et al. Validation of a SNP-based non-invasive prenatal test to detect the fetal 22q11.2 deletion in maternal plasma samples. PLoS One. 2018;13(2):e0193476.
- 40. Santamaria R, Bermejo B, Cigarrán S, Benn P. A national referral laboratory's experience with the implementation of SNP-based non-invasive prenatal screening for fetal aneuploidy and select microdele-tion syndromes. J Fetal Med. 2018;5(1):7–12.
- Van Opstal D, Van Maarle MC, Lichtenbelt K, Weiss MM, Schuring-Blom H, Bhola SL, et al. Origin and clinical relevance of chromosomal aberrations other than the common trisomies detected by genome-wide NIPS: results of the TRIDENT study. Genet Med. 2018;20(5):480-5.
- 42. Chen Y, Yu Q, Mao X, Lei W, He M, Lu W. Noninvasive prenatal testing for chromosome aneuploidies and subchromosomal microdeletions/microduplications in a cohort of 42,910 single pregnancies with different clinical features. Hum Genomics. 2019;13(1):1–8.
- 43. de Wergifosse S, Bevilacqua E, Mezela I, El Haddad S, Gounongbe C, de Marchin J, et al. Cell-free DNA analysis in maternal blood: comparing genome-wide versus targeted approach as a first-line screening test. J Matern Fetal Neonatal Med. 2019;34(21):3552–61.
- 44. Flowers N, Giouzeppos O, Shi G, Love C, Tsegay A, Harrington T, et al. Clinical utility of non-invasive prenatal testing (NIPT) for the detection of segmental aneuploidy. Abstracts for the 42nd Human Genetics Society of Australasia Annual Scientific Meeting Sydney, New South Wales August 4–7, 2018: plenaries and orals. Twin Res Hum Genet. 2019;21(5):407.
- 45. Guy C, Haji-Sheikhi F, Rowland CM, Anderson B, Owen R, Lacbawan FL, et al. Prenatal cell-free DNA screening for fetal aneuploidy in pregnant women at average or high risk: results from a large US clinical laboratory. Mol Genet Genomic Med. 2019;7(3):e545.
- 46. Hu H, Wang L, Wu J, Zhou P, Fu J, Sun J, et al. Noninvasive prenatal testing for chromosome aneuploidies and subchromosomal microdeletions/microduplications in a cohort of 8141 single pregnancies. Hum Genomics. 2019;13(1):1–9.
- Liang D, Cram DS, Tan H, Linpeng S, Liu Y, Sun H, et al. Clinical utility of noninvasive prenatal screening for expanded chromosome disease syndromes. Genet Med. 2019;21(9):1998–2006.
- 48. Sekelska M, Izsakova A, Kubosova K, Tilandyova P, Csekes E, Kuchova Z, et al. P01.86B detection and validation of subchromosomal aberrations detected as part of routine noninvasive prenatal testing. Abstracts from the 52(nd) European Society of Human Genetics (ESHG) conference: posters. Eur J Hum Genet. 2019;27(Suppl 2):1210–1.

BJOG An International Journal of Obstetrics and Gynaecology

- 49. Yu D, Zhang K, Han M, Pan W, Chen Y, Wang Y, et al. Noninvasive prenatal testing for fetal subchromosomal copy number variations and chromosomal aneuploidy by low-pass whole-genome sequencing. Mol Genet Genomic Med. 2019;7(6):e674.
- Chen Y-S, Wu Y-Q, Zhang Y, Ying C-M. Clinical performance of cellfree fetal DNA testing for fetal aneuploidies and subchromosomal deletions/duplications in a cohort of 19,531 pregnancies. Reprod Dev Med. 2020;4(3):163–8.
- Kleinfinger P, Lohmann L, Luscan A, Trost D, Bidat L, Debarge V, et al. Strategy for use of genome-wide non-invasive prenatal testing for rare autosomal aneuploidies and unbalanced structural chromosomal anomalies. J Clin Med. 2020;9(8):2466.
- Liu Y, Liu H, He Y, Xu W, Ma Q, He Y, et al. Clinical performance of non-invasive prenatal served as a first-tier screening test for trisomy 21, 18, 13 and sex chromosome aneuploidy in a pilot city in China. Hum Genomics. 2020;14(1):1–9.
- 53. Luo Y, Hu H, Jiang L, Ma Y, Zhang R, Xu J, et al. A retrospective analysis the clinic data and follow-up of non-invasive prenatal test in detection of fetal chromosomal aneuploidy in more than 40,000 cases in a single prenatal diagnosis center. Eur J Med Genet. 2020;63(9):104001.
- 54. Oneda B, Sirleto P, Baldinger R, Taralczak M, Joset P, Zweier M, et al. Genome-wide non-invasive prenatal testing in single-and multiplepregnancies at any risk: identification of maternal polymorphisms to reduce the number of unnecessary invasive confirmation testing. Eur J Obstet Gynecol Reprod Biol. 2020;252:19–29.
- 55. Pei Y, Hu L, Liu J, Wen L, Luo X, Lu J, et al. Efficiency of noninvasive prenatal testing for the detection of fetal microdeletions and microduplications in autosomal chromosomes. Mol Genet Genomic Med. 2020;8(8):e1339.
- Togneri FS, Allen SK, Mann K, Holgado E, Morgan S. Cytogenomic results following high-chance non-invasive prenatal testing: a UK national audit. Genet Res. 2020;102:e7.
- 57. Wu X, Li Y, Xie X, Su L, Cai M, Lin N, et al. Clinical review of noninvasive prenatal testing: experience from 551 pregnancies with noninvasive prenatal testing-positive results in a tertiary referral center. J Mol Diagn. 2020;22(12):1469–75.
- Xu J, Xue Y, Wang J, Zhou Q, Zhang B, Yu B, et al. The necessity of prenatal diagnosis by CMA for the women with NIPS-positive results. Int J Genomics. 2020;2020:2145701.
- Yin L, Tang Y, Lu Q, Pan A, Shi M. Application value of NIPT for uncommon fetal chromosomal abnormalities. Mol Cytogenet. 2020;13(1):1–7.
- Bevilacqua E, Jani J, Chaoui R, Suk EK, Palma-Dias R, Ko TM, et al. Performance of a targeted cell-free DNA prenatal test for 22q11. 2 deletion in a large clinical cohort. Ultrasound Obstet Gynecol. 2021;58(4):597–602.
- 61. Bu X, Zeng L, Li H, Zhou S, Hu L, He J. Value of chromosomal microarray analysis for the prenatal diagnosis of pregnancy with high risk signaled by non-invasive prenatal testing. Zhonghua Yi Xue Yi Chuan Xue Za Zhi. 2021;38(6):541–4.
- 62. Chen S, Zhang L, Gao J, Li S, Chang C, Chen Y, et al. Expanding the scope of non-invasive prenatal testing to detect fetal chromosomal copy number variations. Front Mol Biosci. 2021;8:350.
- Chen Y, Lai Y, Xu F, Qin H, Tang Y, Huang X, et al. The application of expanded noninvasive prenatal screening for genome-wide chromosomal abnormalities and genetic counseling. J Matern Fetal Neonatal Med. 2021;34(16):2710–6.
- 64. Ge Y, Li J, Zhuang J, Zhang J, Huang Y, Tan M, et al. Expanded noninvasive prenatal testing for fetal aneuploidy and copy number variations and parental willingness for invasive diagnosis in a cohort of 18,516 cases. BMC Med Genomics. 2021;14(1):1–12.
- 65. Gou L, Suo F, Wang Y, Wang N, Wu Q, Hu S, et al. Clinical value for the detection of fetal chromosomal deletions/duplications by noninvasive prenatal testing in clinical practice. Mol Genet Genomic Med. 2021;9(6):e1687.
- 66. Hui L, Loughry L, Halliday J. The expanding scope of noninvasive prenatal testing (NIPT) in Victoria: a population-based study

of prenatal diagnosis after NIPT. Abstracts for the human genetics Society of Australasia Virtual Conference 24–25 November 2020. Twin Res Hum Genet. 2021;24(1):87–8.

- 67. Lai Y, Zhu X, He S, Dong Z, Tang Y, Xu F, et al. Performance of cellfree DNA screening for fetal common aneuploidies and sex chromosomal abnormalities: a prospective study from a less developed autonomous region in Mainland China. Genes. 2021;12(4):478.
- Lin T-Y, Cheng P-J, Hung T-H, Chan K-S, Tsai C, Shaw SW. Taiwanese clinical experience with noninvasive prenatal testing for DiGeorge syndrome. Fetal Diagn Ther. 2021;48(9):672–7.
- Mitchell L, Orefice R, Robertson M, editors. Audit of genotypic outcome in pregnancies screened positive for 22q deletion with cell-free DNA testing. Aust N Z J Obstet Gynaecol. 2021;61(S1):11.
- Pang Y, Wang C, Tang J, Zhu J. Clinical application of noninvasive prenatal testing in the detection of fetal chromosomal diseases. Mol Cytogenet. 2021;14(1):1–11.
- Rafalko J, Soster E, Caldwell S, Almasri E, Westover T, Weinblatt V, et al. Genome-wide cell-free DNA screening: a focus on copy-number variants. Genet Med. 2021;23(10):1847–53.
- Shi P, Wang Y, Liang H, Hou Y, Chen D, Zhao G, et al. The potential of expanded noninvasive prenatal screening for detection of microdeletion and microduplication syndromes. Prenat Diagn. 2021;41(10):1332-42.
- Soster E, Boomer T, Hicks S, Caldwell S, Dyr B, Chibuk J, et al. Three years of clinical experience with a genome-wide cfDNA screening test for aneuploidies and copy-number variants. Genet Med. 2021;23(7):1349–55.
- 74. Wang C, Tang J, Tong K, Huang D, Tu H, Li Q, et al. Expanding the application of non-invasive prenatal testing in the detection of foetal chromosomal copy number variations. BMC Med Genomics. 2021;14(1):1–11.
- Zhu X, Chen M, Wang H, Guo Y, Chau M, Yan H, et al. Clinical utility of expanded non-invasive prenatal screening and chromosomal microarray analysis in high-risk pregnancy. Ultrasound Obstet Gynecol. 2021;57(3):459–65.
- Dar P, Jacobsson B, Clifton R, Egbert M, Malone F, Wapner RJ, et al. Cell-free DNA screening for prenatal detection of 22q11.2 deletion syndrome. Am J Obstet Gynecol. 2022;227(1):79.e1–11.
- 77. Dar P, MacPherson C, Jacobsson B, Egbert M, Malone FD, Wapner RJ, et al. cfDNA prenatal screening for Cri-Du-Chat, Prader-Willi/ Angelman and 1p36del syndromes in 10,971 pregnancies with genetic confirmation. Am J Obstet Gynecol. 2022;226(1):S526–S7.
- Hu Y, Liu W, He G, Xu J, Peng Y, Wang J. Clinical utility of expanded NIPT for chromosomal abnormalities and etiology analysis of cytogenetic discrepancies cases. J Assist Reprod Genet. 2022;39(1):267–79.
- Scott F, Menezes M, Smet M, Carey K, Hardy T, Fullston T, et al. Influence of fibroids on cell-free DNA screening accuracy. Ultrasound Obstet Gynecol. 2022;59(1):114–9.
- Chen Y, Lu L, Zhang Y, Wang F, Ni Y, Wang Q, et al. Clinical application of expanded noninvasive prenatal testing for fetal chromosome abnormalities in a cohort of 39,580 pregnancies. Am J Med Genet A. 2022;188(5):1426–34.
- Raymond Y, Fernando S, Menezes M, Meagher S, Mol BW, McLennan A, et al. Cell-free DNA screening for rare autosomal trisomies and segmental chromosome imbalances. Prenat Diagn. 2022;42:1349–57.
- 82. Yang L, Bu G, Ma Y, Zhao J, Rezak J, La X. Comparison of noninvasive prenatal screening for defined pathogenic microdeletion/microduplication syndromes and nonsyndromic copy number variations: a large multicenter study. J Comp Eff Res. 2022;11:1277–91.
- Zheng Y, Li J, Zhang J, Yang H. The accuracy and feasibility of noninvasive prenatal testing in a consecutive series of 20,626 pregnancies with different clinical characteristics. J Clin Lab Anal. 2022;36:e24660.
- 84. Chen YS, Wang FF, Lu LKY, Ni YH, Wang Q, Ying CM. [Clinical application of expanded noninvasive prenatal testing for fetal chromosome abnormalities]. Zhonghua Yu Fang Yi Xue Za Zhi. 2021;55(12):1491–5.

14710528, 2023, 6, Downloaded from https://obgyn.onlinelibrary.wiley.com/doi/10.1111/1471-0528.17386 by University Of Aberdeen, Wiley Online Library on [23:08/2023]. See the Terms and Conditions (https://onlinelibrary.wiley. and ns) on Wiley Online Library for rules of use; OA articles are governed by the applicable Creative Commons

558

- Dai R, Yu Y, Zhang H, Li L, Jiang Y, Liu R, et al. Analysis of 17,428 pregnant women undergoing non-invasive prenatal testing for fetal chromosome in Northeast China. Medicine (Baltimore). 2021;100(6):e24740.
 - Olenev AS, Baranova EE, Sagaydak OV, Galaktionova AM, Kuznetsova ES, Kaplanova MT, et al. Adoption of a non-invasive prenatal test (NIPT) in prenatal screening in Moscow: first results. Russ Open Medical J. 2021;10(1):110.
 - Pertile MD, Flowers N, Vavrek D, Andrews D, Kalista T, Craig A, et al. Performance of a paired-end sequencing-based noninvasive prenatal screening test in the detection of genome-wide fetal chromosomal anomalies. Clin Chem. 2021;67(9):1210–9.
 - Wang J-W, Lyu Y-N, Qiao B, Li Y, Zhang Y, Dhanyamraju PK, et al. Cell-free fetal DNA testing and its correlation with prenatal indications. BMC Pregnancy Childbirth. 2021;21(1):1–9.
 - Wardrop JCS, Boomer T, Cacheris P, McCullough R. Genome wide non-invasive prenatal testing – 2,000 samples outcome experience. Poster abstracts of the ISPD 21st international conference on prenatal diagnosis and therapy, San Diego, California, USA, 9-12 July 2017. Prenat Diagn. 2017;37(S1):21–105.
 - 90. Chau MHK, Cao Y, Kwok YKY, Chan S, Chan YM, Wang H, et al. Characteristics and mode of inheritance of pathogenic copy number

variants in prenatal diagnosis. Am J Obstet Gynecol. 2019;221(5):493. e1-11.

 Kobelka C, Mattman A, Langlois S. An evaluation of the decisionmaking process regarding amniocentesis following a screen-positive maternal serum screen result. Prenat Diagn. 2009;29(5):514–9.

SUPPORTING INFORMATION

Additional supporting information can be found online in the Supporting Information section at the end of this article.

How to cite this article: Raymond YC, Acreman ML, Bussolaro S, Mol BW, Fernando S, Menezes M, et al. The accuracy of cell-free DNA screening for fetal segmental copy number variants: A systematic review and meta-analysis. BJOG. 2023;130(6):549–559. <u>https://doi.org/10.1111/1471-0528.17386</u>