

# SCIENTIFIC REPORTS



OPEN

## Plasma non-esterified docosahexaenoic acid is the major pool supplying the brain

Chuck T. Chen<sup>1</sup>, Alex P. Kitson<sup>1</sup>, Kathryn E. Hopperton<sup>1</sup>, Anthony F. Domenichiello<sup>1</sup>, Marc-Olivier Trépanier<sup>2</sup>, Lauren E. Lin<sup>3</sup>, Leonardo Ermini<sup>2</sup>, Martin Post<sup>2</sup>, Frank Thies<sup>3</sup> & Richard P. Bazinet<sup>1</sup>

Received: 17 June 2015

Accepted: 05 October 2015

Published: 29 October 2015

Despite being critical for normal brain function, the pools that supply docosahexaenoic acid (DHA) to the brain are not agreed upon. Using multiple kinetic models in free-living adult rats, we first demonstrate that DHA uptake from the plasma non-esterified fatty acid (NEFA) pool predicts brain uptake of DHA upon oral administration, which enters the plasma NEFA pool as well as multiple plasma esterified pools. The rate of DHA loss by the brain is similar to the uptake from the plasma NEFA pool. Furthermore, upon acute iv administration, although more radiolabeled lysophosphatidylcholine (LPC)-DHA enters the brain than NEFA-DHA, this is due to the longer plasma half-life and exposure to the brain. Direct comparison of the uptake rate of LPC-DHA and NEFA-DHA demonstrates that uptake of NEFA-DHA into the brain is 10-fold greater than LPC-DHA. In conclusion, plasma NEFA-DHA is the major plasma pool supplying the brain.

The brain has a unique fatty acid composition, being relatively enriched in certain saturated, monounsaturated and polyunsaturated fatty acids (PUFA), while very low in others. Within the brain, docosahexaenoic acid (DHA; 22:6n-3) is the major omega-3 (n-3) PUFA comprising about 10–15% of all fatty acids and over 50% of all brain PUFA<sup>1</sup>. DHA is a precursor to a series of bioactive molecules coined specialized pro-resolving mediators which includes the resolvins, protectins and maresins as well as docosahexaenoyl ethanolamide<sup>2,3</sup>. Collectively, DHA and its enzymatically synthesized products regulate numerous processes within the brain, including neurogenesis, cell survival, neuroinflammation and its resolution<sup>2,4–9</sup>. Altered DHA signaling has been implicated in a variety of neurological disorders and psychiatric diseases and is currently an active area of research for brain therapeutics<sup>10–12</sup>.

DHA cannot be synthesized *de novo* and must be obtained directly in the diet or via the conversion of shorter chain essential n-3 PUFA, including  $\alpha$ -linolenic acid, to DHA predominately in the liver<sup>13,14</sup>. As the brain synthesis of DHA is very low, it is generally agreed that the brain is supplied with DHA from the plasma<sup>15</sup>. In adults, the concentration of DHA in the brain is at steady-state<sup>8,16</sup>, with the uptake rate closely approximating the consumption rate. Because of this, the rate of brain DHA uptake can be estimated from the rate of brain DHA loss.

In plasma, DHA is present in two major pools: 1) bound to albumin as non-esterified DHA (NEFA-DHA) or lysophosphatidylcholine-esterified DHA (LPC-DHA), or 2) in lipoproteins esterified to triacylglycerol (TAG), diacylglycerol (DAG), cholesteryl ester (CE), phospholipids or other lysophospholipids. There has been considerable controversy as to which plasma fatty acid pools supply the brain with DHA. DHA within lipoproteins, NEFA-DHA and LPC-DHA have all been proposed not only to enter the brain, but to be the major pool supplying the brain with DHA<sup>17</sup>. Plasma lipoproteins account

<sup>1</sup>Department of Nutritional Sciences, Faculty of Medicine, University of Toronto, FitzGerald Building, 150 College St., Room 306, Toronto, ON, Canada, M5S 3E2. <sup>2</sup>Program in Physiology and Experimental Medicine, Peter Gilgan Centre for Research and Learning, The Hospital for Sick Children, Toronto, ON, Canada, M5G 0A4. <sup>3</sup>School of Medicine and Dentistry, University of Aberdeen, Foresterhill, Aberdeen, UK. Correspondence and requests for materials should be addressed to R.P.B. (email: richard.bazinet@utoronto.ca)

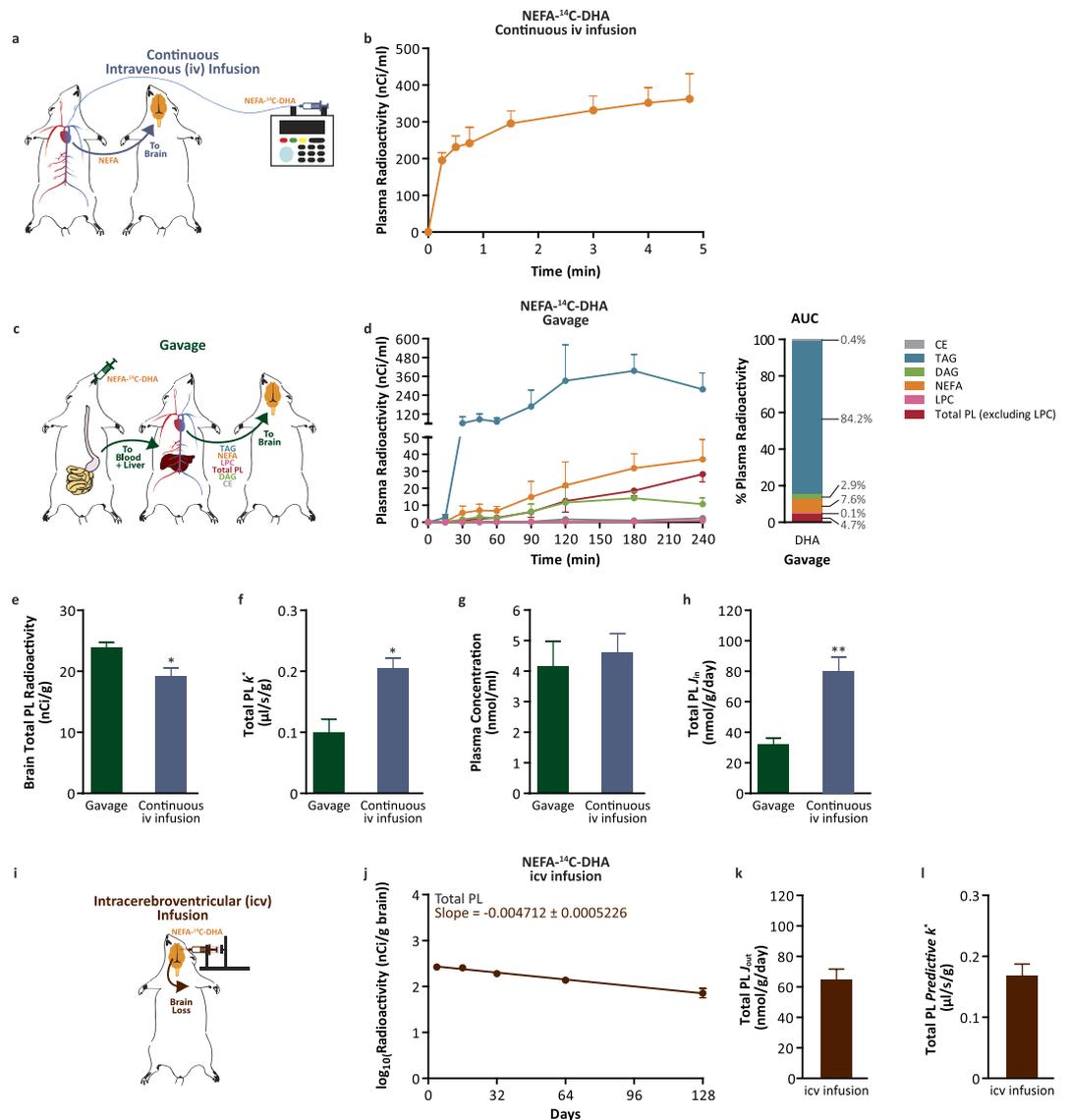
for the largest plasma pool of DHA; however, lipoprotein receptors do not transport cholesterol into the brain and LDL and VLDL receptor knockouts demonstrated that these receptors are not necessary for maintaining brain DHA levels<sup>17,18</sup>. With regards to the plasma NEFA-DHA pool, the observation that the rate of NEFA-DHA uptake into the brain closely matches the rate of brain DHA consumption support the hypothesis that plasma NEFA-DHA is the major source of DHA for the brain<sup>13,16</sup>. However, there is also evidence that LPC-DHA can supply DHA for the brain, as radioactive brain DHA levels are higher upon intravenous (iv) infusion of radiolabeled LPC-DHA compared to NEFA-DHA in developing rats, where DHA accretion is substantial<sup>19–21</sup>. This finding was recently confirmed in adult mice and interpreted as LPC-DHA being the major plasma pool supplying the brain with DHA<sup>22</sup>. However, this conclusion suggests that much more DHA is entering the brain than is being consumed, which is inconsistent with the finding that there is minimal, if any, accretion of DHA in the adult brain<sup>16</sup>. Therefore, to test the contribution of various DHA containing plasma pools to brain DHA uptake we used a series of kinetic approaches that account for the amount of exposure of the brain to the radiotracer and the specific activity of the plasma lipid pool<sup>23,24</sup>. We measured the uptake rate of iv infused NEFA-DHA into the brain, brain DHA uptake upon oral administration which could occur from multiple plasma pools, and the rate of brain DHA consumption. After accounting for an experimentally derived brain DHA uptake rate from NEFA-DHA, the brain uptake from esterified DHA (including LPC-DHA) was negligible, and uptake from NEFA-DHA was similar to the brain DHA consumption rate. We then reproduced the finding that iv infusion of radiolabeled LPC-DHA leads to higher brain radioactivity compared to NEFA-DHA; however, upon correcting for brain exposure and/or the plasma pool size, NEFA-DHA was determined to be the major plasma pool supplying the brain. In our last experiment, we continuously infused NEFA-DHA and LPC-DHA and found that the uptake rate of plasma NEFA-DHA into the brain was 10-fold higher than LPC-DHA.

## Results

**Brain DHA concentration and distribution.** Similar to other reports, the brain total phospholipid DHA concentration of rats fed a chow containing 5%  $\alpha$ -linolenate as the only n-3 PUFA was  $5956 \pm 477$  nmol/g brain (Supplementary Fig. 1a)<sup>25</sup>. Upon further separation of the major structural phospholipid classes, DHA was highly enriched in ethanolamine glycerophospholipids (EtnGpl) ( $4826 \pm 558$  nmol/g brain) as compared to choline glycerophospholipids (ChoGpl) ( $852 \pm 57$  nmol/g brain); whereas between the major signaling phospholipids, DHA was found mostly esterified to phosphatidylserine (PtdSer) ( $1676 \pm 302$  nmol/g brain) as compared to phosphatidylinositol (PtdIns) ( $75 \pm 18$  nmol/g brain). DHA-containing phospholipid species were ubiquitous across brain regions (Supplementary Fig. 1b,c).

**Brain DHA uptake.** To examine the rate of uptake of NEFA-DHA into the brain, albumin-bound NEFA-<sup>14</sup>C-DHA was continuously infused into the jugular vein of rats over a 5-minute period while blood samples were collected from the carotid artery (Fig. 1a). The 5-minute continuous infusion captured the appearance of NEFA-<sup>14</sup>C-DHA in the plasma (Fig. 1b). Measures for the uptake coefficient ( $k^*$ ) and rate of NEFA-DHA incorporation ( $J_{in}$ ) were  $0.20 \pm 0.02$   $\mu$ l/s/g and  $80 \pm 10$  nmol/g/day, respectively (Eqs 4 and 5). To determine the relative contribution of esterified and NEFA-DHA to total brain DHA uptake, NEFA-<sup>14</sup>C-DHA was gavaged (Fig. 1c) to rats resulting in the appearance of <sup>14</sup>C-DHA in several plasma esterified pools as well as the plasma NEFA pool (Fig. 1d; Supplementary Fig. 2 for confirmation of tracer identity). Similar to others, we observed that DHA made up greater than 99% of the radioactivity<sup>16</sup>). By continuous sampling from the carotid artery, it was determined that <sup>14</sup>C-DHA appeared largely in triacylglycerol (TAG; 84% of total plasma radiotracer) and relatively lower amounts were found esterified in phospholipids (total phospholipids excluding LPC; 4.7%), diacylglycerol (DAG; 2.9%), cholesteryl esters (CE; 0.4%) and lysophosphatidylcholine (LPC; 0.13%). NEFA-<sup>14</sup>C-DHA was the second most abundant radiolabeled lipid pool in the plasma after NEFA-<sup>14</sup>C-DHA gavage accounting for 7.6% of total plasma radiotracer area under the curve (Fig. 1d). Radiolabeled LPC-DHA did not appear in the plasma until 3 hours post-ingestion of NEFA-<sup>14</sup>C-DHA which is consistent with it being secreted from the liver and not upon intestinal absorption (Fig. 1d). Using the measured  $k^*$  for NEFA-DHA (Fig. 1f), it was determined that the uptake of esterified DHA ( $k^*_{ES}$ ) was  $-0.0089 \pm 0.002$   $\mu$ l/s/g suggesting that esterified DHA including LPC-DHA is not, quantitatively, a major source for supplying the brain with DHA (Equations 1–3; Supplementary Table 1 for individual phospholipid species). Therefore, if it is assumed that only the radiolabeled DHA in the plasma NEFA pool post-gavage enters the brain, this leads to a  $k^*$  of  $0.098 \pm 0.05$   $\mu$ l/s/g and a net rate of incorporation ( $J_{in}$ ) of  $32 \pm 9$  nmol/g/day (Fig. 1f,h Eqs 4 and 5). Collectively, we find that the  $k^*$  and  $J_{in}$  upon a 5-min continuous infusion are slightly higher than would be predicted from the plasma NEFA pool upon oral gavage of the NEFA-<sup>14</sup>C-DHA tracer (Fig. 1f,g). One reason for this might be error in estimating the AUC between the time points over a 4 hour study and due to acute catabolism of the tracer upon its entry into the brain during the 4 hours which has been estimated to be as much as 20% for PUFA<sup>23</sup>. While the mechanism for the decrease in PUFA radioactivity is not clear, it is possible that a proportion of the tracer enters an unstable pool within the brain.

To examine whether these rates of brain DHA uptake were similar to rates of DHA loss, the rate of loss of DHA from brain phospholipids was determined by administering NEFA-<sup>14</sup>C-DHA

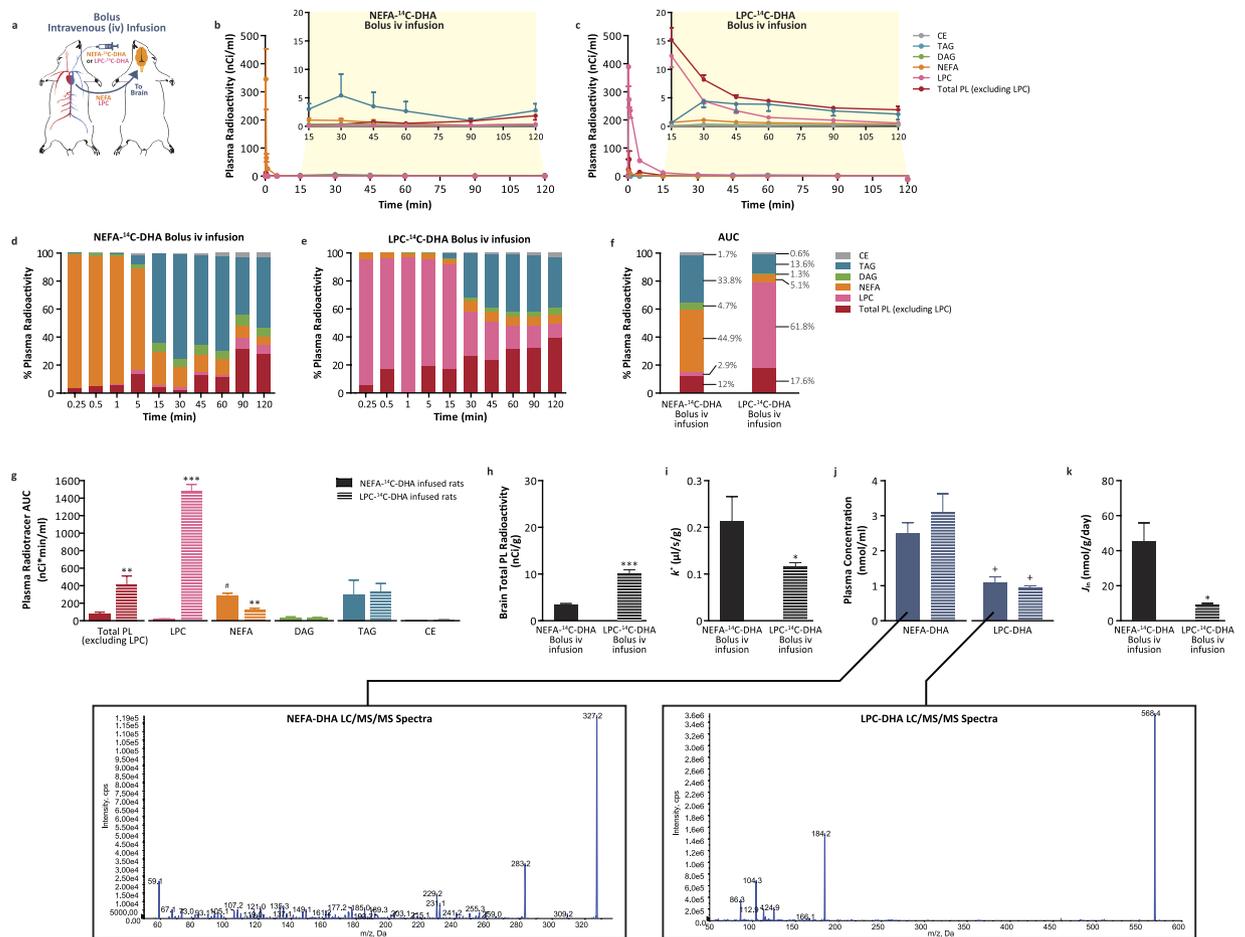


**Figure 1. NEFA-DHA uptake by the brain.** (a) Schematic diagram of kinetic modeling following continuous intravenous (iv) infusion. (b) Plasma lipid radioactivity upon continuous iv infusion of 86  $\mu\text{Ci}$  NEFA-<sup>14</sup>C-DHA over 5 minutes ( $n = 6$ ). Radioactivity from other plasma lipid pools was below detection, 23 pCi. (c) Schematic diagram of kinetic modeling following NEFA-<sup>14</sup>C-DHA gavage. (d) Plasma lipid radioactivity upon NEFA-<sup>14</sup>C-DHA oral gavage over 4 hours ( $n = 4$ ) and the percent composition of total radiolabeled lipid pools in the plasma post-gavage of NEFA-<sup>14</sup>C-DHA. (e) Brain radioactivity of total phospholipids from rats gavaged or continuously iv infused with NEFA-<sup>14</sup>C-DHA. (f) The uptake incorporation coefficient of NEFA-<sup>14</sup>C-DHA ( $k^*$ ) into brain total phospholipids from rat gavage or continuous iv infusion. (g) The plasma concentration of NEFA-DHA prior to NEFA-<sup>14</sup>C-DHA gavage or continuous iv infusion. (h) The net incorporation rate of DHA ( $J_{in}$ ) to brain total phospholipids from rats gavaged or iv infused with NEFA-<sup>14</sup>C-DHA. (i) Schematic diagram of kinetic modeling following intracerebroventricular (icv) infusion. (j) Brain loss curve of esterified <sup>14</sup>C-DHA from brain total phospholipids including lysophosphatidylcholine over 128 days ( $n = 3-4$  independent samples per time point). (k) The loss of DHA from brain total phospholipids (rate of loss,  $J_{out}$ ) from rats infused icv with NEFA-<sup>14</sup>C-DHA which was not significantly different from the  $J_{in}$  of rats continuously iv infused with NEFA-<sup>14</sup>C-DHA. (l) Calculated predictive  $k^*$  from the NEFA-<sup>14</sup>C-DHA icv infusion which was not significantly different from  $k^*$  from NEFA-<sup>14</sup>C-DHA iv infusion. All data are expressed as mean  $\pm$  SEM; whereas percent plasma radioactivity data are expressed as mean. \* $P < 0.05$  and \*\* $P < 0.01$  indicate significant difference from gavage using Students'  $t$ -test. AUC, area under the curve; CE, cholesteryl ester; DAG, diacylglycerol; LPC, lysophosphatidylcholine; NEFA, non-esterified fatty acid; PL, phospholipids; TAG, triacylglycerol.

intracerebroventricularly and calculating the slope of DHA loss from total phospholipids (Fig. 1i, Supplementary Fig. 3 for individual phospholipid species). This measured loss half-life ( $t_{1/2}$ ) of  $64 \pm 7$  days (Equation 6) translated to rate of loss from brain phospholipids ( $J_{out}$ ) of  $65 \pm 7$  nmol/g/day (Fig. 1k, Equation 7) and a predictive  $k^*$  of  $0.17 \pm 0.02 \mu\text{l/s/g}$  (Fig. 1l, Equation 8). The  $t_{1/2}$  and  $J_{out}$  are in broad agreement with previous reports by DeMar *et al.* and Lin *et al.*, with minor discrepancies possibly related to differences to animal ages, diets and strain<sup>16,26</sup>. Therefore, it is important that comparisons of fatty acid kinetic parameters from various models be from studies conducted under similar conditions. The  $k^*$  and  $J_{in}$  from the continuous iv infusion are not significantly different from the predictive  $k^*$  and  $J_{out}$  again suggesting that plasma NEFA-DHA can replace brain DHA loss to maintain homeostatic levels of brain DHA. The results predict that plasma NEFA-DHA is the major contributor to brain DHA *in vivo*.

**The relative partitioning of LPC-DHA into the brain compared with NEFA-DHA.** Earlier work demonstrated that upon iv infusion of a bolus of radiolabeled NEFA-DHA or LPC-DHA, that LPC-DHA preferentially targeted the brain<sup>19,20</sup>. Recently, with the discovery of a putative LPC-DHA transporter, and using similar experiments, it was concluded that LPC-DHA represents the major mechanism by which DHA enters the brain<sup>22</sup>. However, the results presented above (Fig. 1), are not compatible with the hypothesis that LPC-DHA is the major plasma pool supplying the brain. Thus, we set out to reproduce the finding of higher brain radioactivity following iv LPC-<sup>14</sup>C-DHA infusion. This time, however, the amount of DHA exposure to the brain was measured, which was not considered in the previous report. Similar to other reports<sup>19,22</sup>, a bolus iv infusion of radiolabeled NEFA-DHA or LPC-DHA was used and brains were collected 2 hours post-infusion (Fig. 2a). Consistent with the short half-life of plasma NEFA of approximately 30 seconds, NEFA-<sup>14</sup>C-DHA decreased 207-fold from the peak initial level within 5 minutes (Fig. 2b). In contrast, radiolabeled LPC-DHA was still elevated at 5 minutes and had only decreased 7-fold from the peak. Furthermore, at 30 minutes, the reduction in plasma LPC-<sup>14</sup>C-DHA levels was merely 88-fold as compared to a 339-fold reduction in plasma NEFA-<sup>14</sup>C-DHA levels, suggesting a much longer half-life for LPC-DHA (Fig. 2c). This translated to higher total relative LPC-<sup>14</sup>C-DHA exposure (61.8% of total plasma radiotracer) in rats receiving LPC-<sup>14</sup>C-DHA as compared to NEFA-<sup>14</sup>C-DHA exposure (44.9% of total plasma radiotracer) in rats receiving NEFA-<sup>14</sup>C-DHA (Fig. 2d–f). Collectively, the AUC for NEFA-DHA in the iv NEFA-<sup>14</sup>C-DHA infused rats was  $282 \pm 32$  nCi\*min/ml, while the AUC for plasma LPC-DHA in the iv LPC-<sup>14</sup>C-DHA infused rats was  $1478 \pm 80$  nCi\*min/ml (Fig. 2g). Similar to other reports, we found that radiolabeled brain DHA was 3-fold higher in iv LPC-<sup>14</sup>C-DHA infused animals compared those receiving NEFA-<sup>14</sup>C-DHA (Fig. 2h). However, because the radiolabel appeared in multiple plasma pools over the 2 hours, it is unclear which plasma pool would have supplied the brain with DHA. Nevertheless, we corrected brain radioactivity for either plasma LPC-<sup>14</sup>C-DHA or NEFA-<sup>14</sup>C-DHA exposure, which was 5.2 fold higher for LPC-DHA than NEFA-DHA (Fig. 2g). An uptake coefficient ( $k^*$ ) can be calculated, which was significantly higher for NEFA-DHA ( $0.21 \pm 0.05 \mu\text{l/s/g}$ ) than LPC-DHA ( $0.12 \pm 0.009 \mu\text{l/s/g}$ ) (Fig. 2i). Finally, to determine the rate of brain DHA uptake, the concentration of plasma NEFA-DHA and LPC-DHA was measured using LC/MS/MS and found that the concentration of LPC-DHA was 64% lower than NEFA-DHA (Fig. 2j). Applying the plasma concentration to the  $k^*$  for LPC-DHA and NEFA-DHA, the net rate of entry ( $J_{in}$ ) into the brain from the plasma NEFA pool ( $46 \pm 10$  nmol/g/day) is 5-fold higher than the rate of entry from the plasma LPC-DHA pool ( $9 \pm 0.6$  nmol/g/day) (Fig. 2k). Because a small fraction of LPC-DHA is in the form of lipoproteins, this method slightly overestimates albumin-bound plasma LPC-DHA. However, similar to the gavage experiment, if other pools are entering the brain, these rates could be overestimates. Since the  $k^*$  from the current bolus iv NEFA-DHA infusion study is similar to our results with continuous iv NEFA infusion, it suggests that, at least for this pool, it is not an overestimate.

To circumvent the repackaging of infused radiotracer into multiple plasma pools and to control for exposure, the uptake of NEFA-<sup>14</sup>C-DHA and LPC-<sup>14</sup>C-DHA was quantified using the continuous 5-minute infusion model (Fig. 3a). Consistent with the shorter half-life of NEFA-DHA, the plasma radioactivity appeared to plateau around 1 min (Fig. 3b), while plasma radioactivity increased during the study for the LPC-DHA (Fig. 3c). At the 5-minute time point, 97% of the total lipid radioactivity in the <sup>14</sup>C-DHA-infused rats was in the plasma NEFA pool and 97% of the total lipid radioactivity in LPC-<sup>14</sup>C-DHA-infused rats was in the plasma phospholipid pool. As predicted from the previous experiments, brain radioactivity was significantly higher upon 5-minute LPC-<sup>14</sup>C-DHA infusion (Fig. 3d). After correction for plasma exposure, however, the incorporation coefficient ( $k^*$ ) was significantly higher in the group receiving NEFA-<sup>14</sup>C-DHA ( $0.18 \pm 0.04 \mu\text{l/s/g}$ ) than LPC-<sup>14</sup>C-DHA ( $0.06 \pm 0.009 \mu\text{l/s/g}$ ) (Fig. 3e). The elevated  $k^*$  in the <sup>14</sup>C-DHA-infused rats was magnified upon correction for the plasma pool sizes (Fig. 3f) as plasma NEFA-DHA pool ( $J_{in} = 46 \pm 11$  nmol/g/day) contributes 10-fold more DHA than plasma LPC-DHA ( $J_{in} = 4.7 \pm 1.2$  nmol/g/day) (Fig. 3g). Importantly plasma concentrations of NEFA-DHA and LPC-DHA is this and the previous experiments are similar to the normal physiological range reported in the literature<sup>27,28</sup>. The replacement half-life of DHA in brain total phospholipids would be  $65 \pm 12$  days from the plasma NEFA-DHA pool and  $676 \pm 122$  days from the plasma LPC-DHA pool (calculated based on the concentrations from Supplementary Fig. 1). Furthermore, if use the experimentally derived  $k^*$  (Fig. 3e) for LPC-DHA to predict the amount of brain radioactive DHA upon the oral gavage of NEFA-<sup>14</sup>C-DHA (Fig. 1e), this leads to a 148 fold underestimate of the measured radioactivity. The incorporation coefficients and net rate of entry for NEFA-DHA were consistent across

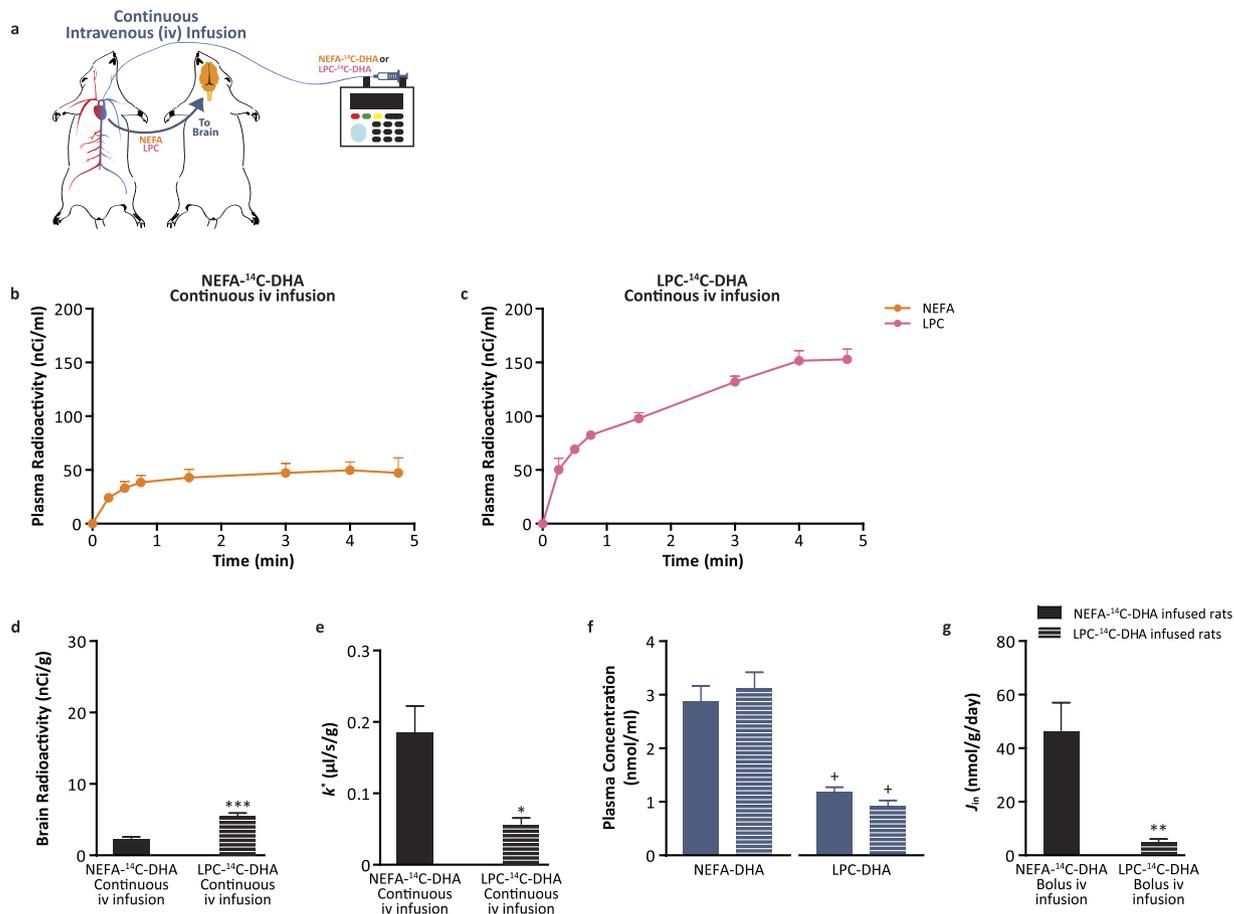


**Figure 2. Plasma LPC-DHA contribution to brain total DHA uptake via bolus iv infusion kinetic modeling.** (a) Schematic diagram of iv infusion kinetic modeling. (b,c) Plasma lipid radioactivity from NEFA-<sup>14</sup>C-DHA or LPC-<sup>14</sup>C-DHA bolus iv infusion (13 μCi) followed by 2-hour plasma sampling (n = 4 each). (d,e) Percent composition of various radiolabeled lipid pools in the plasma at each time point following an acute iv bolus infusion of NEFA-<sup>14</sup>C-DHA or LPC-<sup>14</sup>C-DHA. (f) Percent composition of various radiolabeled lipid pools in the plasma over 2-hour period following an acute iv bolus infusion of NEFA-<sup>14</sup>C-DHA or LPC-<sup>14</sup>C-DHA. (g) Plasma radiotracer exposure from various lipid pools after bolus iv infusion of NEFA-<sup>14</sup>C-DHA or LPC-<sup>14</sup>C-DHA. (h) Brain radioactivity of total phospholipids from rats iv infused with bolus NEFA-<sup>14</sup>C-DHA or LPC-<sup>14</sup>C-DHA. (i) The uptake incorporation coefficient ( $k'$ ) into brain total phospholipids from rats iv infused with bolus NEFA-<sup>14</sup>C-DHA or LPC-<sup>14</sup>C-DHA. (j) The baseline plasma concentration of total NEFA-DHA and LPC-DHA prior to NEFA-<sup>14</sup>C-DHA or LPC-<sup>14</sup>C-DHA bolus iv infusion. LC/MS/MS spectra of NEFA-DHA and LPC-DHA. (k) The net incorporation rate of DHA ( $J_m$ ) into brain total phospholipids from rats iv infused with bolus NEFA-<sup>14</sup>C-DHA or LPC-<sup>14</sup>C-DHA. Percent plasma radioactivity data are expressed as mean. Remaining data are expressed as mean ± SEM. \* $P < 0.05$ , \*\* $P < 0.01$  and \*\*\* $P < 0.001$  indicate significant difference from NEFA-<sup>14</sup>C-DHA infused rats. + $P < 0.01$  indicates significant difference between plasma LPC-DHA and NEFA-DHA concentration. # $P < 0.001$  indicates significant difference between radiolabeled LPC pool from LPC-<sup>14</sup>C-DHA infused rats and radiolabeled NEFA-DHA pool from NEFA-<sup>14</sup>C-DHA infused rats. Students'  $t$ -test was used to determine significant difference. See Fig. 2 for abbreviations.

all studies for the plasma NEFA-DHA, despite differences in time and dose. This is not surprising as uptake of plasma NEFA-DHA is thought to be via passive diffusion and the models used in this study are time-independent<sup>29–33</sup>.

## Discussion

Using several kinetic models, these studies demonstrated that plasma NEFA-DHA is the major plasma pool supplying the brain with DHA in free-living adult rats. The finding that plasma NEFA-DHA is the major plasma pool supplying the brain does not negate earlier important findings of preferential LPC-DHA incorporation, as compared to NEFA-DHA, into the brain<sup>19,20,22</sup>. Similar to the reports of



**Figure 3. Plasma LPC-DHA and NEFA-DHA contribution to brain total DHA uptake via continuous iv infusion kinetic model.** (a) Schematic diagram of continuous iv infusion kinetic modeling. (b,c) Plasma curve from continuous iv infusion of low-dose (10  $\mu$ Ci) NEFA-<sup>14</sup>C-DHA or LPC-<sup>14</sup>C-DHA over 5 minutes ( $n = 4$  each). (d) Brain radioactivity of total phospholipids from rats iv infused with NEFA-<sup>14</sup>C-DHA or LPC-<sup>14</sup>C-DHA. (e) The uptake incorporation coefficient ( $k'$ ) into brain total phospholipids from rats continuously iv infused with NEFA-<sup>14</sup>C-DHA or LPC-<sup>14</sup>C-DHA. (f) The baseline plasma concentration of total NEFA-DHA and LPC-DHA prior to NEFA-<sup>14</sup>C-DHA or LPC-<sup>14</sup>C-DHA continuous iv infusion. (g) The net incorporation rate of DHA ( $J_{in}$ ) into brain total phospholipids from rats continuously iv infused with NEFA-<sup>14</sup>C-DHA or LPC-<sup>14</sup>C-DHA. All data are expressed as mean  $\pm$  SEM. \* $P < 0.05$ , \*\* $P < 0.01$  and \*\*\* $P < 0.001$  indicate significant difference from NEFA-<sup>14</sup>C-DHA infused rats using Students'  $t$ -test. + $P < 0.01$  indicates significant difference between plasma LPC-DHA and NEFA-DHA concentration. See Fig. 2 for abbreviations.

others<sup>20,22,34</sup>, our results suggest that upon a single intravenous bolus dose or infusion of LPC-DHA, more LPC-derived DHA will target the brain as compared to NEFA-DHA. This has therapeutic importance for studies aiming to deliver DHA to the brain and is consistent with preclinical findings regarding the efficacy of LPC-DHA analogues targeting the brain in models of stroke<sup>35</sup>. However, despite this “preferential LPC-DHA uptake”, LPC-DHA is not the major mechanism by which DHA enters the brain under normal conditions. LPC-DHA appears to circulate with a longer plasma half-life than NEFA-DHA. When controlling for plasma exposure and/or the steady-state non-labeled “cold” pool size, plasma NEFA-DHA is the major pool supplying the brain with DHA *in vivo*. It is important to note, that while the NEFA and LPC-DHA tracers disappear from the plasma with different half-lives, the cold concentrations remain at steady-state due to replacement from other pools. Based on results from experiments similar to ours (Fig. 2) and because mice lacking the candidate transporter for LPC-DHA *Mfsd2a* had deficits in brain DHA levels (~8% in phospholipids) compared to wild-type controls (~20%), Nguyen and colleagues proposed that the major mechanism by which DHA enters the brain is from plasma-derived LPC<sup>22</sup>. The large decrease in brain phospholipid DHA in *Mfsd2a* knockout mice may appear to be at odds with our finding that LPC-DHA is a relatively minor source of brain DHA, but there are many possible explanations for this apparent discrepancy, including: (a) the low DHA levels in the *Mfsd2a* knockout are due to the large loss of DHA-rich neuronal cells in the knockout (over 50% loss of Purkinje cells), possibly

also explaining why DHA supplementation does not restore brain DHA levels or (b) that the *Mfsd2a* knockout was life-long and much brain DHA accretion occurs during development which may require LPC-DHA. Furthermore it is worth noting that despite life-long deficiency of *Mfsd2a*, knockout mice still have 8% DHA which is similar to what is reported during omega-3 deficiency in the mouse, albeit methodological differences may account for this result<sup>36,37</sup>.

The finding that plasma NEFA is the major plasma pool supplying the brain with DHA suggests that the previous studies that have estimated brain DHA uptake to be between 2.4 and 3.8 mg/brain/day, by imaging brain DHA uptake upon infusion of labeled NEFA-DHA in humans, are only slight underestimates of the total brain DHA uptake rate<sup>7,38,39</sup>. While it is known that the plasma NEFA pool is supplied by the adipose and circulating lipoproteins<sup>40</sup>, studies on the source of DHA in the plasma NEFA pool are, to the best of our knowledge, lacking. The DHA content in rat white adipose tissue was recently reported to be 13 mg<sup>41</sup>. Given our calculated rate of uptake of about 20 µg/day, this would suggest that adipose could supply for the brain for 1.8 years. However, this is an overestimate as DHA released from adipose does not solely supply to the brain and released DHA would be taken up by other tissues. Even though the brain/body partition coefficient for DHA has not been reported for rats, the partition coefficient has been estimated to be 5% for palmitate in rats and 0.5% for DHA in a monkey<sup>23,39</sup>. It is important to note that these might also be underestimates as fatty acids that are taken up by other tissues may be released and taken up by the brain at a later time. Also, a proportion, if not all, of the released DHA from adipose is being replaced from the diet<sup>4</sup>. Using similar calculations, we have recently estimated that adipose in adult humans contains enough DHA (20–50 g) to supply the brain for 14–36 years<sup>42</sup>.

In summary, our results demonstrate that the NEFA pool of DHA is the major source of DHA supplying the rat brain *in vivo* and confirm that a bolus of plasma LPC-DHA will target the brain more than a bolus of NEFA-DHA. Collectively, our results resolve a more than 20 year-old controversy and will inform studies aiming to deliver or image/quantify DHA entry into the brain which is being examined in stroke<sup>43,44</sup>, traumatic brain injury<sup>8,45</sup> and other neurological diseases<sup>12,38,46,47</sup>. Future studies should examine the effects of diet, genetics, brain injury, disease, development and aging to test if the mechanisms presented here persist.

## Online Methods

All procedures were performed in accordance with the policies set out by the Canadian Council on Animal Care and were approved by the Animal Ethics Committee at the University of Toronto. Male Sprague Dawley rats were purchased from Charles Rivers (Saint-Constant, QC, Canada) at 10 weeks of age and kept at the animal facility with automated 12 hour light-dark cycle and a constant temperature of 22 °C for five weeks. The rats received *ad libitum* access to water and standard chow (Teklad 2018, Harlan, Madison, WI, USA) which was composed of 54% of linoleate (18:2n-6), 5% of  $\alpha$ -linolenate (18:3n-3) and <0.3% of longer chained PUFA (20:2n-6, 20:3n-3, 20:4n-6, 20:5n-3, 22:4n-6, 22:5n-6, 22:5n-3 and DHA), as measured by gas chromatography-flame ionization detection (GC-FID).

**Radiotracer perfusate.** Radiolabeled NEFA-<sup>14</sup>C-DHA ([1-<sup>14</sup>C]-DHA, specific activity: 58 mCi/mmol, Moravek Biochemical Inc., Brea, CA, USA) were solubilized in olive oil for gavage, and 5 mM HEPES buffer containing 50 mg/ml fatty acid-free bovine serum albumin for intravenous (iv) infusion (continuous and bolus) or 5 mM HEPES buffer containing 100 mg/ml fatty acid-free bovine serum albumin for intracerebroventricular (icv) infusion. The NEFA-<sup>14</sup>C-DHA perfusates were prepared to a concentration of 100 µCi/ml, 78 µCi/ml or 2 µCi/µl for the gavage, iv or icv study, respectively. The purity of <sup>14</sup>C-radiotracer was >99.9% as confirmed by high performance liquid chromatography (HPLC) and liquid scintillation counting (LSC) (Supp Fig. 1). Radiolabeled LPC-<sup>14</sup>C-DHA (1-docosahexaenoyl-2-lysophosphocholine (Lysophosphatidylcholine, L-a-1-docosahexaenoyl [1-<sup>14</sup>C]), specific activity: 55 mCi/mmol, American Radiolabeled Chemicals Inc., St. Louis, MO, USA) were solubilized in 5 mM HEPES buffer containing 100 mg/ml fatty acid-free bovine serum albumin for iv infusions. The LPC-<sup>14</sup>C-DHA perfusates were prepared to a concentration of 33 µCi/ml or 8.6 µCi/ml for iv bolus and continuous infusion, respectively.

**Gavage.** Four 15-week-old rats were anesthetised with isofluorane inhalation (3% induction, 1–2% maintenance). Rats were given a lower back subcutaneous injection of 100 mg/kg of Ketoprofen (MERIAL Canada, Inc., Baie d'Urfé, QC, Canada). Polyethylene catheters (PE 50, Intramedic™, Becton Dickinson, Franklin Lakes, NJ, USA) with silicone tubing (Silicone tubing 0.020 in (I.D.) and 0.037 in (O.D.), VWR®, Mississauga, ON, Canada) filled with 0.9% saline were implanted into the carotid artery. Surgery lasted for approximately 15 minutes. After surgery, all rats were singly housed to recover from anesthesia for 24 hours with *ad libitum* access to food and water. Rats were not fasted for the radiotracer oral gavage. Twenty-four hours post-surgery, rats were gavaged with 500 µCi of NEFA-<sup>14</sup>C-DHA (8.62 µmol DHA) in olive oil. Blood samples were obtained prior to gavage and at 15, 30, 45, 60, 90, 120, 180, and 240-minute post-gavage. After 4 hours, the rats were rapidly euthanized by head-focused, high-energy microwave irradiation (MI; 13.5 kW for 1.6 seconds; Cober Electronics Inc., Norwalk, CT, USA). Brains were excised and stored at –80 °C for radioactive and biochemical analyses. Plasma was isolated from whole blood via microcentrifugation at 6200 rpm (2000 g) for 10 minutes and stored at –80 °C.

**Intravenous (iv) continuous tracer infusion.** Fourteen 15-week-old rats were anesthetized with isoflurane inhalation (3% induction, 1–2% maintenance). Rats were implanted with polyethylene catheters into the right carotid artery and right jugular vein that were buried in a skin pouch above the scapula and randomized to receive either an iv dose of 86  $\mu\text{Ci}$  NEFA- $^{14}\text{C}$ -DHA, 10  $\mu\text{Ci}$  NEFA- $^{14}\text{C}$ -DHA or 10  $\mu\text{Ci}$  LPC- $^{14}\text{C}$ -DHA which translated to 1.48  $\mu\text{mol}$ , 0.17  $\mu\text{mol}$  or 0.18  $\mu\text{mol}$  of DHA over 5 minutes as previously described<sup>25</sup>.

**Intravenous (iv) bolus tracer infusion.** Eight 15-week-old rats were implanted with catheters in the right carotid artery and right jugular vein which were externalized and protected by skin button (Braintree Scientific, Braintree, MA, USA) sutured to the subcutaneous tissue at the scapula, and hand-infused with 13  $\mu\text{Ci}$  of either NEFA- $^{14}\text{C}$ -DHA or LPC- $^{14}\text{C}$ -DHA which translated to 0.22  $\mu\text{mol}$  or 0.24  $\mu\text{mol}$  of DHA, respectively. Following hand-infusion of radiotracer, the catheter was immediately flushed with saline. Blood samples were collected at 0, 0.25, 0.5, 1, 5, 15, 30, 45, 60, 90, 120 minutes post-infusion. After two hours, the rats were rapidly euthanized by MI. Brains were excised and stored at  $-80^\circ\text{C}$  for radioactive and biochemical analyses. Plasma was isolated from whole blood via microcentrifugation and stored at  $-80^\circ\text{C}$ .

**Intracerebroventricular (icv) tracer infusion.** At 15 weeks of age, eight rats were euthanized by MI for baseline brain fatty acid concentration measurements and seventeen 15-week-old rats were administered a 5  $\mu\text{l}$  icv injection containing 10  $\mu\text{Ci}$  of  $^{14}\text{C}$ -DHA as previously described by Chen *et al.*<sup>48</sup>.

**Lipid analysis.** Total lipids from whole brain (icv), one brain hemisphere (gavage and iv) and plasma (gavage and iv) were extracted and various neutral lipid and phospholipid classes from total lipid extract were isolated as previously described by Chen *et al.*<sup>48</sup>. Lysophosphatidylcholine (LPC) and lysophosphatidylethanolamine (LPE) from plasma were isolated by the method of Wang and Gustafson<sup>49</sup>. LPC and LPE were separated on TLC H-plates (Analtech, Newark, DE, USA) along with authentic standards. A day prior to sample loading, plates were washed in 0.4% ammonium sulphate. Plates were removed to air dry at room temperature for 2 hours and activated at  $110^\circ\text{C}$  for 20 minutes prior to use. Samples were loaded onto 4 cm lanes. Plates were developed chloroform:methanol:acetic acid:acetone: water (40:25:7:4:2 by vol). Bands corresponding to authentic standards of LPC and LPE were visualized under UV light after spraying with 0.1% 8-anilino-1-naphthalene sulfonic acid and collected for liquid scintillation counting (LSC).

**NEFA-DHA and LPC-DHA analysis.** Liquid chromatography (LC)/tandem mass spectrometry (MS/MS) was performed on an Agilent 1290 HPLC System (Agilent Technologies: Santa Clara, California, USA) and an ABSciex QTRAP5500 Mass Spectrometer (AB Sciex, Framingham, Massachusetts, USA). NEFA-DHA chromatography ran at a flow rate of 600  $\mu\text{L}/\text{min}$  on an Agilent Zorbax SD-Phenyl 3.5  $\mu\text{m}$  3.0  $\times$  50 mm column (Agilent Technologies). The mobile phase consisted of (A) water and (B) acetonitrile, both containing 1  $\mu\text{L}$  propionic acid/250 mL solvent. The gradient was held at initial conditions of 20% B, holding for 2 minutes and ramping to 100% B at 8 minutes, held for 1 minute and returning to initial conditions. The mass spectrometer was operated in negative ESI mode with a source temperature of  $600^\circ\text{C}$ , source voltage of  $-4500\text{ V}$ , curtain gas setting of 30, and collision gas N<sub>2</sub> at medium. Mass transitions used for DHA were 327.15 to 283.15  $m/z$  and for internal standard, DHA-d5 (Cayman Chemical, Ann Arbor, Michigan, USA), is 332.15 to 288.15  $m/z$ . LPC-DHA chromatography ran at a flow rate of 800  $\mu\text{L}/\text{min}$  on a Phenomenex Kinetex HILIC 2.6  $\mu\text{m}$  50  $\times$  4.6 mm column (Phenomenex, Torrance, California, USA). The mobile phase consisted of (A) 90:10 water: acetonitrile and (B) 10:90 water: acetonitrile both containing 5 mM ammonium formate pH 3.2. The gradient was held at initial conditions of 10% A for 1 minute, ramping to 30% A at 3.5 minutes and returning to initial conditions. The mass spectrometer was operated in positive ESI mode with a source temperature of  $600^\circ\text{C}$ , source voltage of  $+5500\text{ V}$ , curtain gas setting of 30, and collision gas N<sub>2</sub> set at medium. Mass transitions used for LPC-DHA were 568.4 to 184.1  $m/z$  and for internal standard, LPC-17:0 (Avanti Polar Lipids), were 510.4 to 184.1  $m/z$ . Data integration and quantitation was performed using AB Sciex Analyst 1.6 software.

**Gas chromatography (GC)-flame ionization detection (FID).** Plasma NEFA concentration and brain fatty acid concentration at baseline were quantified as described by Chen *et al.*<sup>48</sup>.

**High performance liquid chromatography (HPLC)-UV photodiode array detection and liquid scintillation counting (LSC).** Fatty acid methyl esters (FAME) analysis were previously described by Chen *et al.*<sup>48</sup>.

**Matrix-assisted laser desorption/ionization (MALDI)-mass spectral imaging.** The brain tissue block was mounted onto the specimen disc of a cryostat (Leica Microsystems) using optimal cutting temperature (OCT) compound (Sakura Finetek, Torrance, CA). The tissue sections were sliced at a thickness of 12  $\mu\text{m}$  at  $-15^\circ\text{C}$  and mounted onto indium tin oxide (ITO)-coated glass slides. A thin matrix layer was applied to the tissue sections using an automated MALDI plate matrix deposition system (TM-Sprayer<sup>TM</sup>, Leap Technologies, Carrboro, NC). A total of 5 ml of DHB solution (15 mg/ml in 50%

acetonitrile/0.1% trifluoroacetic acid) or 9-aminoacridine (9AA, 15 mg/mL in methanol) was sprayed per slide during 4 passes at 140 °C (DHB) or at 80 °C (9AA) with a velocity of 400 mm/min and a line spacing of 3 mm. A time-of-flight tandem mass spectrometer (AB Sciex TOF/TOFTM 5800 System, AB SCIEX, Ontario, Canada) was used to acquire the images. MALDI-mass spectra were obtained using a Nd:YAG laser (349 nm) at 3 ns pulse width and 400 Hz firing rate. To install the ITO-coated glass slides in the ionization chamber, we used a special holder (AB Sciex) having concavities. The data were acquired in the positive-ion or negative-ion reflector mode using an external calibration method. The external calibration lipids (Avanti Polar Lipids) were deposited on the ITO-coated slides to minimize mass shift. In the imaging experiment, a total of 200 laser shots per point were irradiated (1s/point) and the interval between data points were 80 μm. The mass spectrometric data were processed using a specialized script of Analyst software (AB Sciex) at a mass resolution of 0.1 amu and images were visualized using TissueView software (AB Sciex). After MALDI imaging was completed, the glass coverslip containing the placenta section was carefully removed from the MALDI plate, dipped in methanol to remove the matrix and fixed prior to DAPI staining.

Slices were prepared with Vectashield (Vector Laboratories) mounting medium containing DAPI and visualized at 368nm on a widefield epi-fluorescent microscope at 4× magnification. Multiple images were stitched together to make full image using Nikon Elements Software (NIS-Elements Basic Research 3.10).

**Kinetics.** To estimate the relative contribution of plasma NEFA (UE) and esterified (ES) DHA to brain phospholipids, the following equation were derived:

$$\frac{dC_{brain}^*}{dt} = k_{UE}^* C_{plasma}^* (UE) + k_{ES}^* C_{plasma}^* (ES) \quad (1)$$

where  $C_{brain}^*$  is the radioactivity in the brain (nCi/g brain) from DHA at time,  $T$ ,  $k_{UE/ES}^*$  is unidirectional incorporation coefficient (μl/s/g brain) which represents the incorporation of plasma  $^{14}C$ -DHA into stable brain lipid pools, and  $C_{plasma}^* (UE/ES)$  is the radioactivity in the plasma (nCi/ml plasma) from DHA at time,  $T$ , of NEFA- $^{14}C$ -DHA gavaged rats. Integration of Equation 1 from the start of radiotracer infusion to time of death at  $T$  will yield:

$$C_{brain}^* (T) = k_{UE}^* \int_0^T C_{plasma}^* UE dt + k_{ES}^* \int_0^T C_{plasma}^* ES dt \quad (2)$$

In order to calculate the relative contribution of esterified DHA to brain phospholipids, Equation 2 can be rearranged to solve for  $k_{ES}^*$ :

$$k_{ES}^* = \frac{C_{brain}^* (T) - k_{UE}^* \int_0^T C_{plasma}^* UE dt}{\int_0^T C_{plasma}^* ES dt} \quad (3)$$

However, to calculate  $k_{ES}^*$ ,  $k_{UE}^*$  was separately measured from intravenous infusion of NEFA- $^{14}C$ -DHA where an acute five-minute infusion measures the uptake of albumin-bound NEFA- $^{14}C$ -DHA without  $^{14}C$ -DHA incorporation into phospholipids, triacylglycerol, cholesteryl esters, and diacylglycerol via hepatic lipoprotein packaging.  $k_{UE}^*$  is often referred to as  $k^*$ .

$$k_{UE}^* = \frac{C_{brain}^* (T)}{\int_0^T C_{plasma}^* UE dt} \quad (4)$$

Since the incorporation coefficient applies to both radiolabeled and non-radiolabeled fatty acids, we can further calculate the rate of incorporation of non-radiolabeled plasma DHA into stable brain lipid pools,  $J_{in}$ .

$$J_{in} = k_{UE}^* C_{plasma} \quad (5)$$

where  $C_{plasma}$  is the plasma NEFA-DHA concentration. In addition to the calculation of relative contribution of NEFA-DHA to brain phospholipid DHA uptake, the comparison of  $J_{in}$  from iv infusion and rate of loss,  $J_{out}$ , from icv infusion can infer to the amount of plasma lipid pool(s) contributing to the maintenance of brain phospholipid DHA levels.

Loss half-lives,  $t_{1/2}$  (day), of  $^{14}C$ -DHA in brain total and phospholipid classes were calculated from the slope of regression lines (day $^{-1}$ ) which were obtained by plotting log-transformed  $^{14}C$ -DHA radioactivity against day post-icv infusion.

$$t_{1/2} = \frac{\log_{10} 2}{(\text{slope of regression line})} \quad (6)$$

The  $t_{1/2}$  of DHA was used to calculate the  $J_{out}$  (nmol/g brain/day) from brain phospholipids by the following equation:

$$J_{out} = \frac{0.693C_{brain}}{t_{1/2}} \quad (7)$$

where  $C_{brain}$  is the baseline DHA concentration in a stable phospholipid pool (nmol/g brain). If the NEFA-DHA pool is the major contributor to total brain DHA uptake, then  $J_{in}$  would equal  $J_{out}$ . By rearranging Equation 5, predictive  $k_{UE}^*$  can be solved:

$$\text{Predictive } k_{UE}^* = \frac{J_{out}}{C_{plasma}} \quad (8)$$

Finally, to calculate a fractional loss (%/day) of DHA from stable phospholipid pool.

$$\text{Fractional loss} = \frac{J_{out}}{C_{i,DHA}} * 100 \quad (9)$$

**Statistics.** Concentrations and kinetic parameters are expressed as mean  $\pm$  SEM. HPLC profiles were analyzed as pooled samples and do not have SEM. Statistical comparisons of gavage  $k_{UE}^*/J_{in}$  and iv  $k_{UE}^*/J_{in}$  as well as iv predictive  $k_{UE}^*$  and gavage/iv  $k_{UE}^*$  were performed, *a priori*, using two tailed Student's *t*-test. Statistical significant differences were denoted as \* $p < 0.05$ , \*\* $p < 0.01$  and \*\*\* $p < 0.001$ .

## References

- Diau, G. Y. *et al.* The influence of long chain polyunsaturate supplementation on docosahexaenoic acid and arachidonic acid in baboon neonate central nervous system. *BMC Med* **3**, 11, doi: 10.1186/1741-7015-3-11 (2005).
- Serhan, C. N. Pro-resolving lipid mediators are leads for resolution physiology. *Nature* **510**, 92–101, doi: 10.1038/nature13479 (2014).
- Kim, H. Y., Spector, A. A. & Xiong, Z. M. A synaptogenic amide N-docosahexaenoylethanolamide promotes hippocampal development. *Prostaglandins Other Lipid Mediat* **96**, 114–120, doi: 10.1016/j.prostaglandins.2011.07.002S1098-8823(11)00063-3 (2011).
- Bazinet, R. P. & Laye, S. Polyunsaturated fatty acids and their metabolites in brain function and disease. *Nat Rev Neurosci* **15**, 771–785, doi: 10.1038/nrn3820nrrn3820 (2014).
- Kim, H. Y. Novel metabolism of docosahexaenoic acid in neural cells. *J Biol Chem* **282**, 18661–18665, doi: R70001520010.1074/jbc.R700015200 (2007).
- Liu, J. J., Green, P., John Mann, J., Rapoport, S. I. & Sublette, M. E. Pathways of polyunsaturated fatty acid utilization: Implications for brain function in neuropsychiatric health and disease. *Brain research* **1597C**, 220–246, doi: S0006-8993(14)01666-710.1016/j.brainres.2014.11.059 (2015).
- Cao, D. *et al.* Docosahexaenoic acid promotes hippocampal neuronal development and synaptic function. *J Neurochem* **111**, 510–521, doi: 10.1111/j.1471-4159.2009.06335.xJNC6335 (2009).
- Kim, H. Y. Neuroprotection by docosahexaenoic acid in brain injury. *Military medicine* **179**, 106–111, doi: 10.7202/MILMED-D-14-00162 (2014).
- Calon, F. *et al.* Docosahexaenoic acid protects from dendritic pathology in an Alzheimer's disease mouse model. *Neuron* **43**, 633–645, doi: 10.1016/j.neuron.2004.08.013S0896627304005227 (2004).
- Calon, F. Omega-3 polyunsaturated fatty acids in Alzheimer's disease: key questions and partial answers. *Curr Alzheimer Res* **8**, 470–478, doi: BSP/CAR/0173 (2011).
- Palacios-Pelaez, R., Lukiw, W. J. & Bazan, N. G. Omega-3 essential fatty acids modulate initiation and progression of neurodegenerative disease. *Mol Neurobiol* **41**, 367–374, doi: 10.1007/s12035-010-8139-z (2010).
- Salem, N. Jr., Vandal, M. & Calon, F. The benefit of docosahexaenoic acid for the adult brain in aging and dementia. *Prostaglandins, leukotrienes, and essential fatty acids* **92**, 15–22, doi: 10.1016/j.plefa.2014.10.003 (2015).
- Rapoport, S. L., Rao, J. S. & Igarashi, M. Brain metabolism of nutritionally essential polyunsaturated fatty acids depends on both the diet and the liver. *Prostaglandins Leukot Essent Fatty Acids* **77**, 251–261, doi: S0952-3278(07)00155-X10.1016/j.plefa.2007.10.023 (2007).
- Scott, B. L. & Bazan, N. G. Membrane docosahexaenoate is supplied to the developing brain and retina by the liver. *Proceedings of the National Academy of Sciences of the United States of America* **86**, 2903–2907 (1989).
- DeMar, J. C. Jr., Ma, K., Chang, L., Bell, J. M. & Rapoport, S. I. alpha-Linolenic acid does not contribute appreciably to docosahexaenoic acid within brain phospholipids of adult rats fed a diet enriched in docosahexaenoic acid. *J Neurochem* **94**, 1063–1076 (2005).
- DeMar, J. C. Jr., Ma, K., Bell, J. M. & Rapoport, S. I. Half-lives of docosahexaenoic acid in rat brain phospholipids are prolonged by 15 weeks of nutritional deprivation of n-3 polyunsaturated fatty acids. *Journal of neurochemistry* **91**, 1125–1137, doi: 10.1111/j.1471-4159.2004.02789.x (2004).
- Chen, C. T. & Bazinet, R. P. beta-oxidation and rapid metabolism, but not uptake regulate brain eicosapentaenoic acid levels. *Prostaglandins, leukotrienes, and essential fatty acids* **92**, 33–40, doi: 10.1016/j.plefa.2014.05.007 (2015).
- Dietschy, J. M. Central nervous system: cholesterol turnover, brain development and neurodegeneration. *Biol Chem* **390**, 287–293, doi: 10.1515/BC.2009.035 (2009).
- Thies, F., Pillon, C., Moliere, P., Lagarde, M. & Lecerf, J. Preferential incorporation of sn-2 lysoPC DHA over unesterified DHA in the young rat brain. *The American journal of physiology* **267**, R1273–1279 (1994).
- Thies, F., Delachambre, M. C., Bentejac, M., Lagarde, M. & Lecerf, J. Unsaturated fatty acids esterified in 2-acyl-1-lysophosphatidylcholine bound to albumin are more efficiently taken up by the young rat brain than the unesterified form. *J Neurochem* **59**, 1110–1116 (1992).
- Green, P. & Yavin, E. Mechanisms of docosahexaenoic acid accretion in the fetal brain. *J Neurosci Res* **52**, 129–136, doi: 10.1002/(SICI)1097-4547(19980415)52:2<129::AID-JNR1>3.0.CO;2-C (1998).

22. Nguyen, L. N. *et al.* Mfsd2a is a transporter for the essential omega-3 fatty acid docosahexaenoic acid. *Nature* **509**, 503–506, doi: 10.1038/nature13241 (2014).
23. Robinson, P. J. *et al.* A quantitative method for measuring regional *in vivo* fatty-acid incorporation into and turnover within brain phospholipids: review and critical analysis. *Brain Res Brain Res Rev* **17**, 187–214 (1992).
24. Purdon, D., Arai, T. & Rapoport, S. No evidence for direct incorporation of esterified palmitic acid from plasma into brain lipids of awake adult rat. *J Lipid Res* **38**, 526–530 (1997).
25. Chen, C. T. *et al.* The low levels of eicosapentaenoic acid in rat brain phospholipids are maintained via multiple redundant mechanisms. *J Lipid Res* **54**, 2410–2422, doi: 10.1194/jlr.M038505 (2013).
26. Lin, L. E. *et al.* Chronic dietary n-6 PUFA deprivation leads to conservation of arachidonic acid and more rapid loss of DHA in rat brain phospholipids. *Journal of lipid research* **56**, 390–402, doi: 10.1194/jlr.M055590 (2015).
27. Croset, M., Brossard, N., Polette, A. & Lagarde, M. Characterization of plasma unsaturated lysophosphatidylcholines in human and rat. *Biochem J* **345** Pt 1, 61–67 (2000).
28. Rapoport, S. I., Chang, M. C. & Spector, A. A. Delivery and turnover of plasma-derived essential PUFAs in mammalian brain. *Journal of lipid research* **42**, 678–685 (2001).
29. Kamp, F. & Hamilton, J. A. pH gradients across phospholipid membranes caused by fast flip-flop of un-ionized fatty acids. *Proceedings of the National Academy of Sciences of the United States of America* **89**, 11367–11370 (1992).
30. Kamp, F., Zakim, D., Zhang, F., Noy, N. & Hamilton, J. A. Fatty acid flip-flop in phospholipid bilayers is extremely fast. *Biochemistry* **34**, 11928–11937 (1995).
31. Ouellet, M. *et al.* Diffusion of docosahexaenoic and eicosapentaenoic acids through the blood-brain barrier: An *in situ* cerebral perfusion study. *Neurochemistry international* **55**, 476–482 (2009).
32. Kamp, F. & Hamilton, J. A. How fatty acids of different chain length enter and leave cells by free diffusion. *Prostaglandins Leukot Essent Fatty Acids* **75**, 149–159, doi: S0952-3278(06)00081-010.1016/j.plefa.2006.05.003 (2006).
33. Hamilton, J. A., Guo, W. & Kamp, F. Mechanism of cellular uptake of long-chain fatty acids: Do we need cellular proteins? *Molecular and cellular biochemistry* **239**, 17–23 (2002).
34. Lemaitre-Delaunay, D. *et al.* Blood compartmental metabolism of docosahexaenoic acid (DHA) in humans after ingestion of a single dose of [(13)C]DHA in phosphatidylcholine. *J Lipid Res* **40**, 1867–1874 (1999).
35. Lagarde, M. *et al.* Biological properties of a DHA-containing structured phospholipid (AceDoPC) to target the brain. *Prostaglandins Leukot Essent Fatty Acids* **92**, 63–65, doi: 10.1016/j.plefa.2014.01.005S0952-3278(14)00019-2 (2015).
36. Hussein, N. *et al.* Artificial rearing of infant mice leads to n-3 fatty acid deficiency in cardiac, neural and peripheral tissues. *Lipids* **44**, 685–702, doi: 10.1007/s11745-009-3318-2 (2009).
37. Lafourcade, M. *et al.* Nutritional omega-3 deficiency abolishes endocannabinoid-mediated neuronal functions. *Nat Neurosci* **14**, 345–350, doi: 10.1038/nn.2736 (2011).
38. Umhau, J. C. *et al.* Brain docosahexaenoic acid [DHA] incorporation and blood flow are increased in chronic alcoholics: a positron emission tomography study corrected for cerebral atrophy. *PLoS one* **8**, e75333, doi: 10.1371/journal.pone.0075333 (2013).
39. Umhau, J. C. *et al.* Imaging incorporation of circulating docosahexaenoic acid into the human brain using positron emission tomography. *J Lipid Res* **50**, 1259–1268, doi: 10.1194/jlr.M800530-JLR200 (2009).
40. Frayn, K. N., Summers, L. K. & Fielding, B. A. Regulation of the plasma non-esterified fatty acid concentration in the postprandial state. *Proc Nutr Soc* **56**, 713–721, doi: S0029665197000244 (1997).
41. Salem, N. M. *et al.* Distribution of omega-6 and omega-3 polyunsaturated fatty acids in the whole rat body and 25 compartments. *Prostaglandins Leukot Essent Fatty Acids*, doi: S0952-3278(15)00122-210.1016/j.plefa.2015.06.002 (2015).
42. Domenichello, A. F., Kitson, A. P. & Bazinet, R. P. Is docosahexaenoic acid synthesis from alpha-linolenic acid sufficient to supply the adult brain? *Progress in lipid research* **59**, 54–66, doi: 10.1016/j.plipres.2015.04.002S0163-7827(15)00022-3 (2015).
43. Hong, S. H., Belayev, L., Khoutorova, L., Obenaus, A. & Bazan, N. G. Docosahexaenoic acid confers enduring neuroprotection in experimental stroke. *J Neurol Sci* **338**, 135–141, doi: 10.1016/j.jns.2013.12.033S0022-510X(13)03104-3 (2014).
44. Eady, T. N. *et al.* Docosahexaenoic acid complexed to albumin provides neuroprotection after experimental stroke in aged rats. *Neurobiol Dis* **62**, 1–7, doi: 10.1016/j.nbd.2013.09.008S0969-9961(13)00254-4 (2014).
45. Barrett, E. C., McBurney, M. I. & Ciappio, E. D. omega-3 fatty acid supplementation as a potential therapeutic aid for the recovery from mild traumatic brain injury/concussion. *Advances in nutrition* **5**, 268–277, doi: 10.3945/an.113.005280 (2014).
46. Vandal, M. *et al.* Reduction in DHA transport to the brain of mice expressing human APOE4 compared to APOE2. *Journal of neurochemistry* **129**, 516–526, doi: 10.1111/jnc.12640 (2014).
47. Taha, A. Y., Burnham, W. M. & Auvin, S. Polyunsaturated fatty acids and epilepsy. *Epilepsia* **51**, 1348–1358, doi: 10.1111/j.1528-1167.2010.02654.x (2010).
48. Chen, C. T., Liu, Z. & Bazinet, R. P. Rapid de-esterification and loss of eicosapentaenoic acid from rat brain phospholipids: an intracerebroventricular study. *J Neurochem* **116**, 363–373, doi: 10.1111/j.1471-4159.2010.07116.x (2011).
49. Wang, W. Q. & Gustafson, A. One-dimensional thin-layer chromatographic separation of phospholipids and lysophospholipids from tissue lipid extracts. *J Chromatogr* **581**, 139–142 (1992).

## Acknowledgements

This project was funded by a NSERC and CIHR grant to R.P.B. and studentship to C.T.C. R.P.B. holds a Canada Research Chair in Brain Lipid Metabolism. R.P.B. acknowledges support and mass spectrometry equipment and solutions for lipidomics from Sciex. Computer programmable pump software was designed by Dr. Brian Scott. HPLC analyses were performed at the Analytical Facility at the Department of Nutritional Sciences by Dr. Zhen Liu. LC/MS/MS analyses of NEFA-DHA and LPC-DHA and MALDI imaging were performed at the Analytical Facility for Bioactive Molecules (AFBM) with the assistance of Michael Leadley. The AFBM is part of the Centre for the Study of Complex Childhood Diseases (CSCCD) at the Hospital for Sick Children, Toronto, Ontario. CSCCD was supported by Canadian Foundation for Innovation (CFI).

## Author Contributions

C.T.C. planned the study design, conducted the experiments, analyzed the samples, performed the statistical analysis and wrote the first draft of the manuscript. A.P.K. and K.E.H. conducted the experiments and performed TLC. A.F.D., M.T. and L.E.L. conducted the experiments. L.E. and M.P. performed the MALDI imaging and generated brain images. F.T. provided expertise on LPC methodology. R.P.B.

obtained the funding and planned the study design. All authors have contributed to and reviewed the final manuscript.

### Additional Information

**Supplementary information** accompanies this paper at <http://www.nature.com/srep>

**Competing financial interests:** The authors declare no competing financial interests

**How to cite this article:** Chen, C. T. *et al.* Plasma non-esterified docosahexaenoic acid is the major pool supplying the brain. *Sci. Rep.* **5**, 15791; doi: 10.1038/srep15791 (2015).



This work is licensed under a Creative Commons Attribution 4.0 International License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>